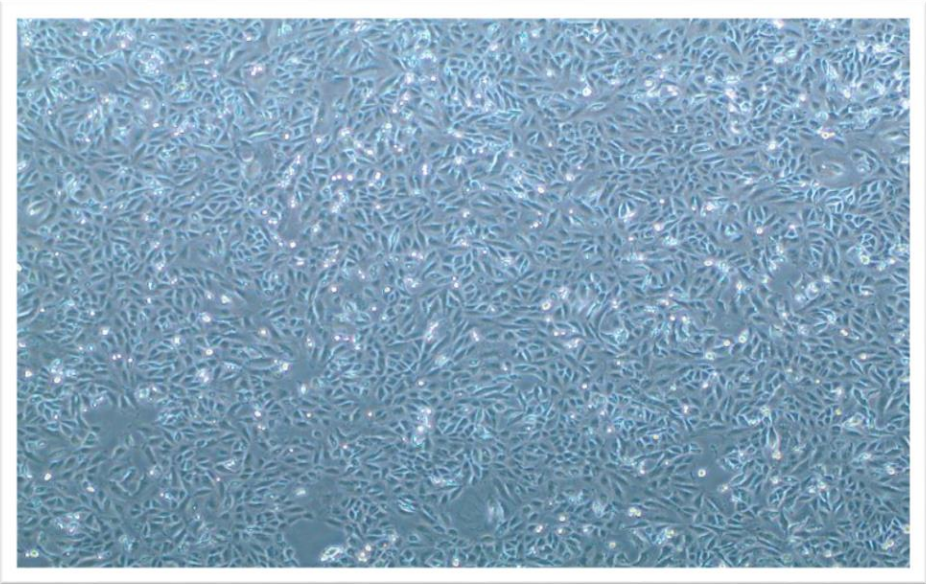


FiberCell Systems Inc.
a better way to grow cells

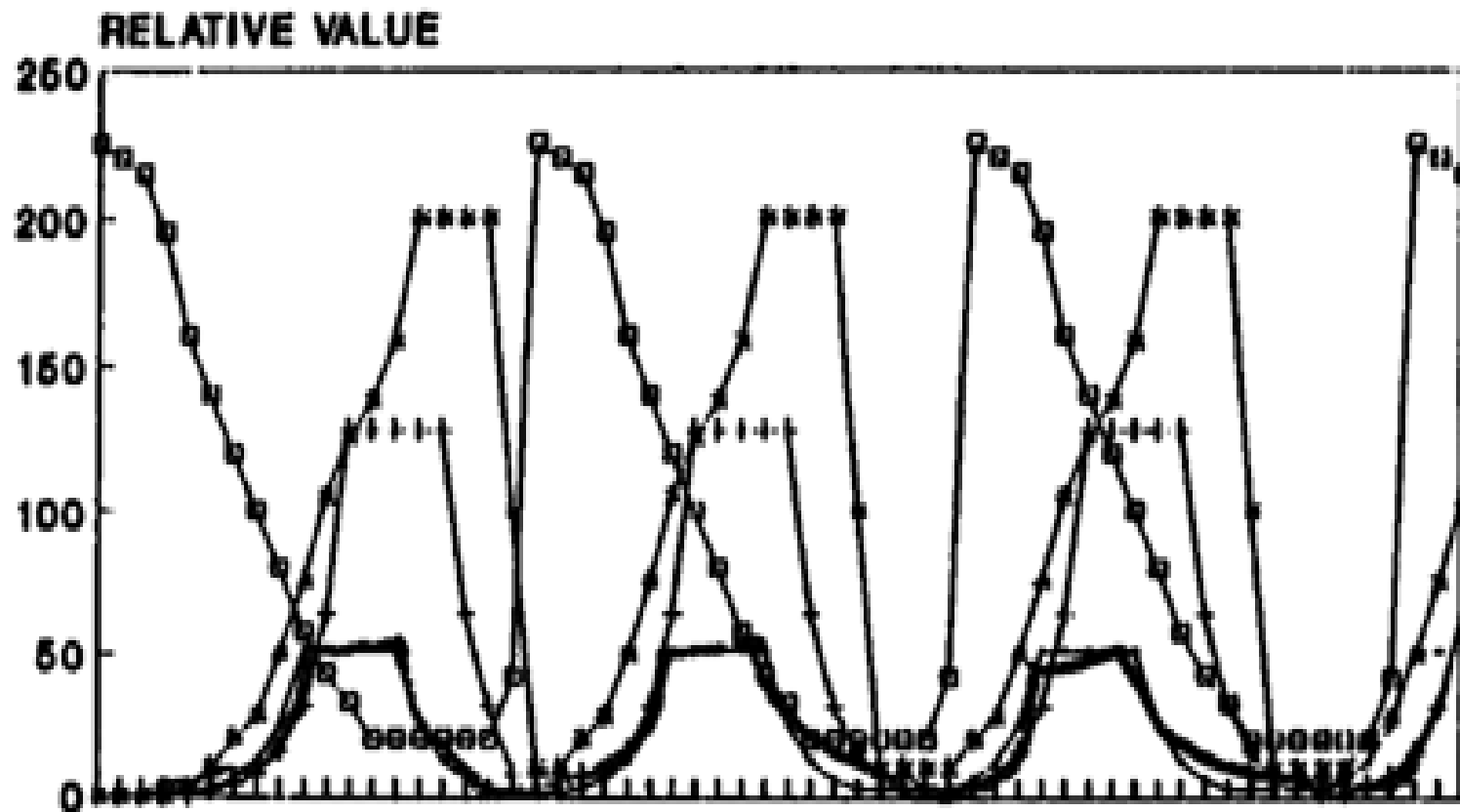
Cell Culture Through the Ages



Cell Culture Options for Scale-Up

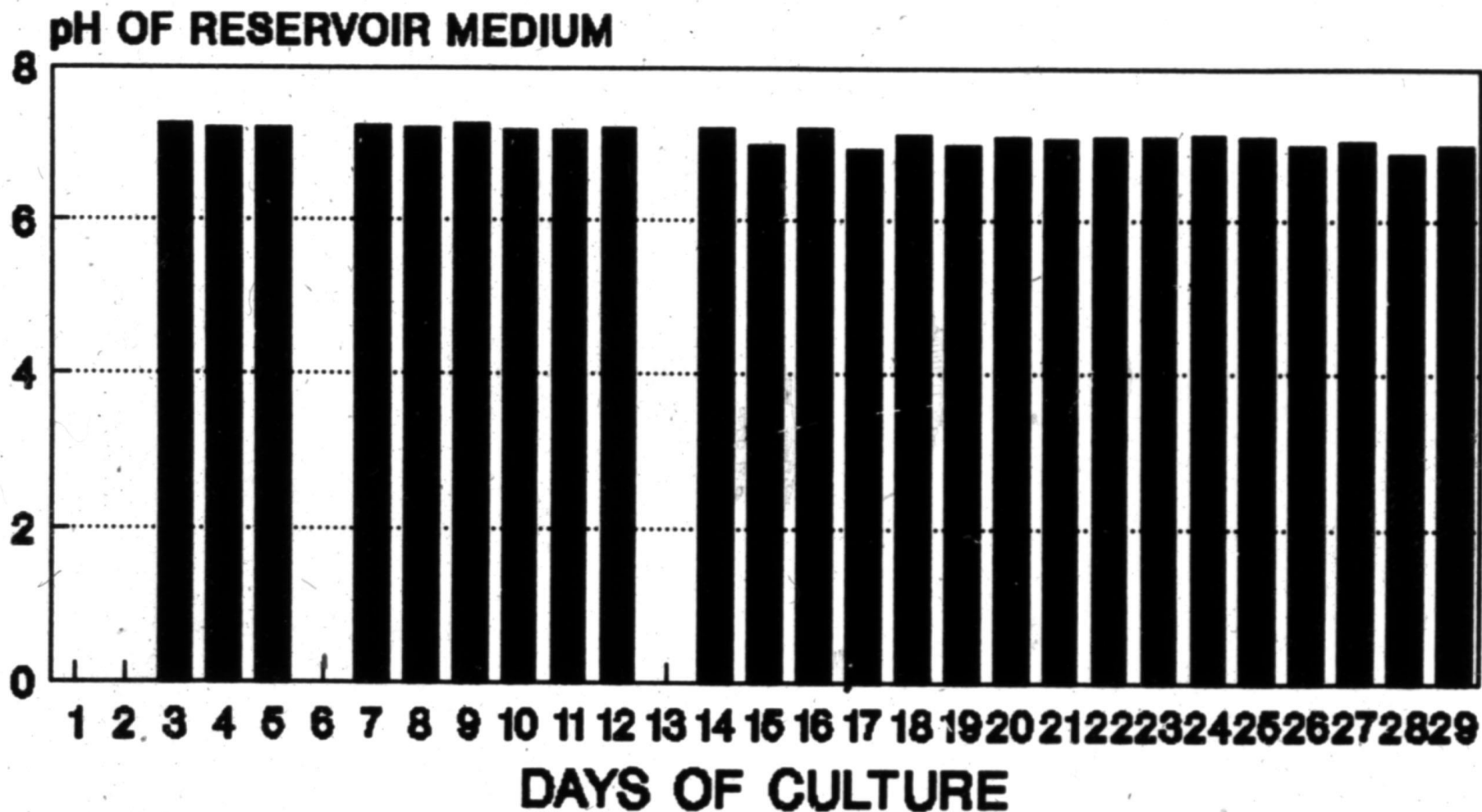
- Roller Bottles
- Cell Factory
- Cell Cube
- Cell Culture Bags
- Spinner Flasks
- Bioreactors

'Feast or Famine'

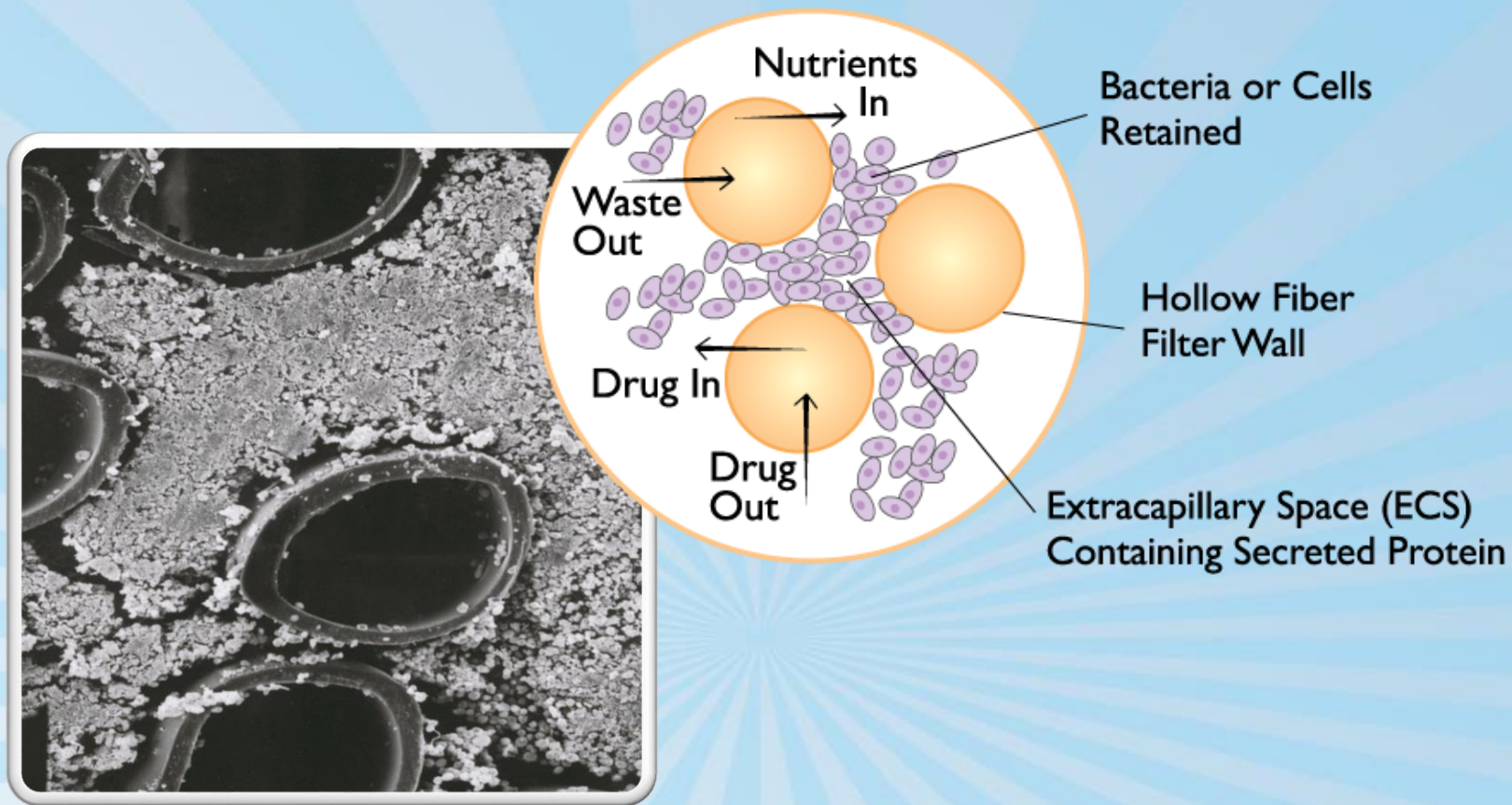


HF CULTURE OF CHO CELLS

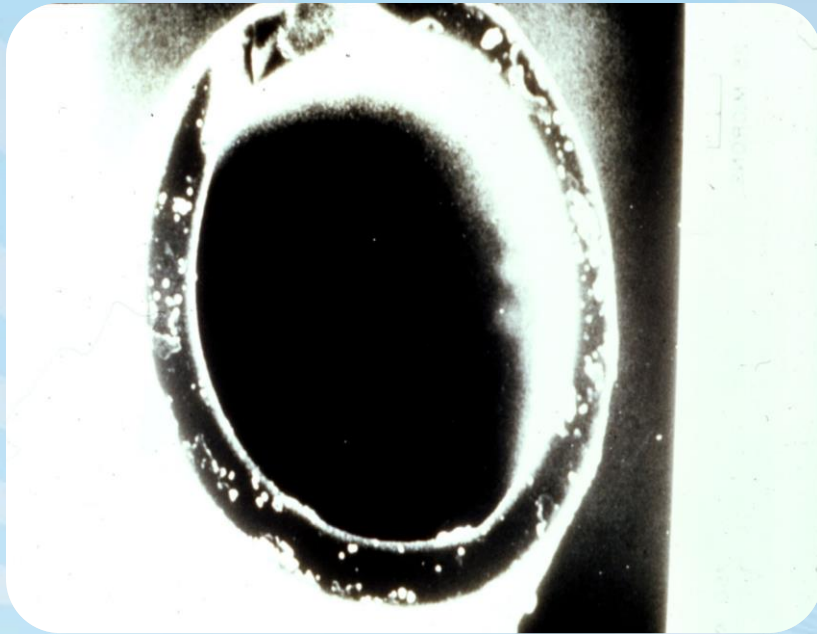
pH CHANGES



Hollow Fiber: How it works

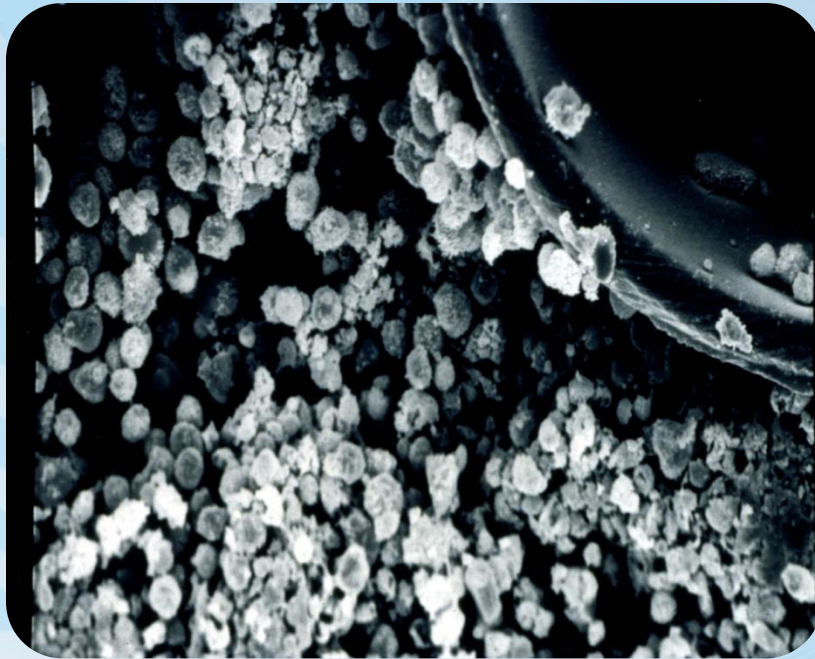


Hollow Fiber Specifications

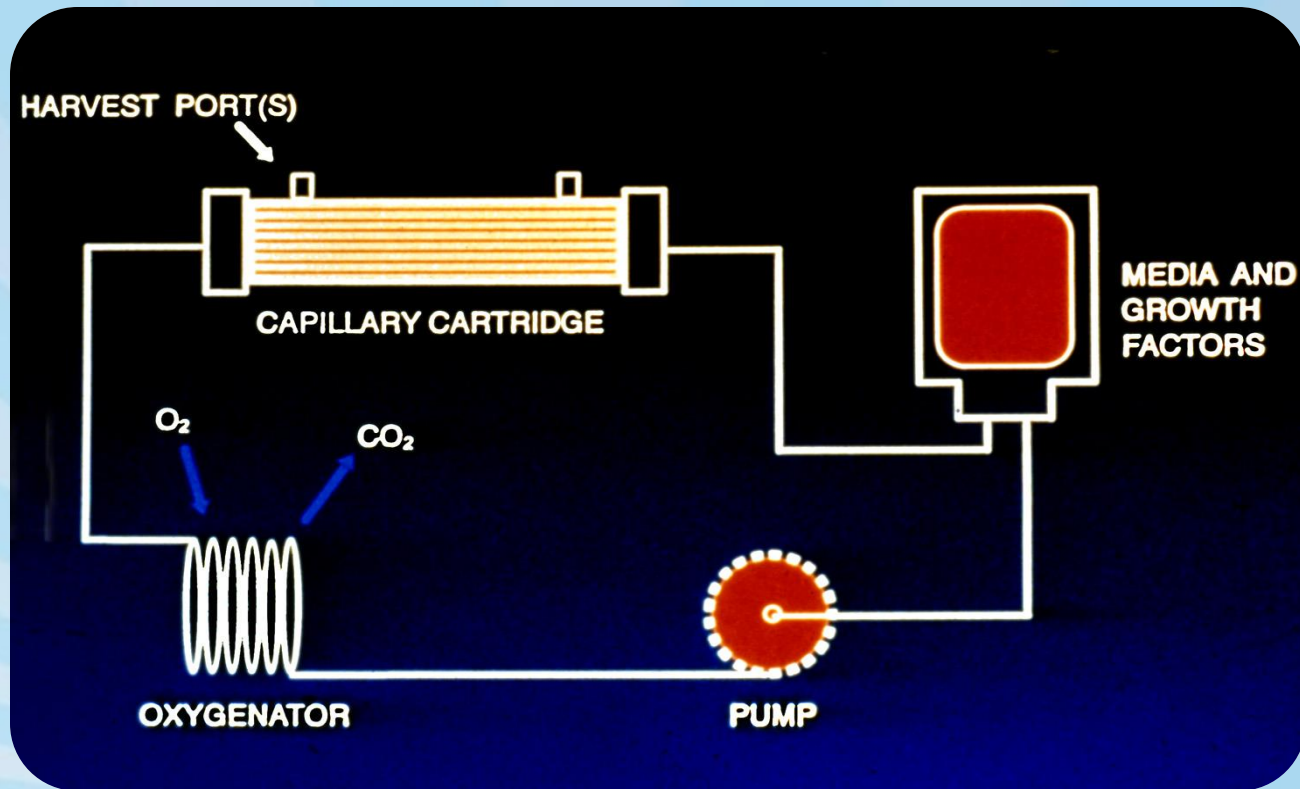


- Hydrophilic Polysulfone or PVDF
- 210 μ m O.D.
- 8 μ m wall thickness
- GFR of 20kd fiber in excess of 140

HF Culture of Lymphocytes

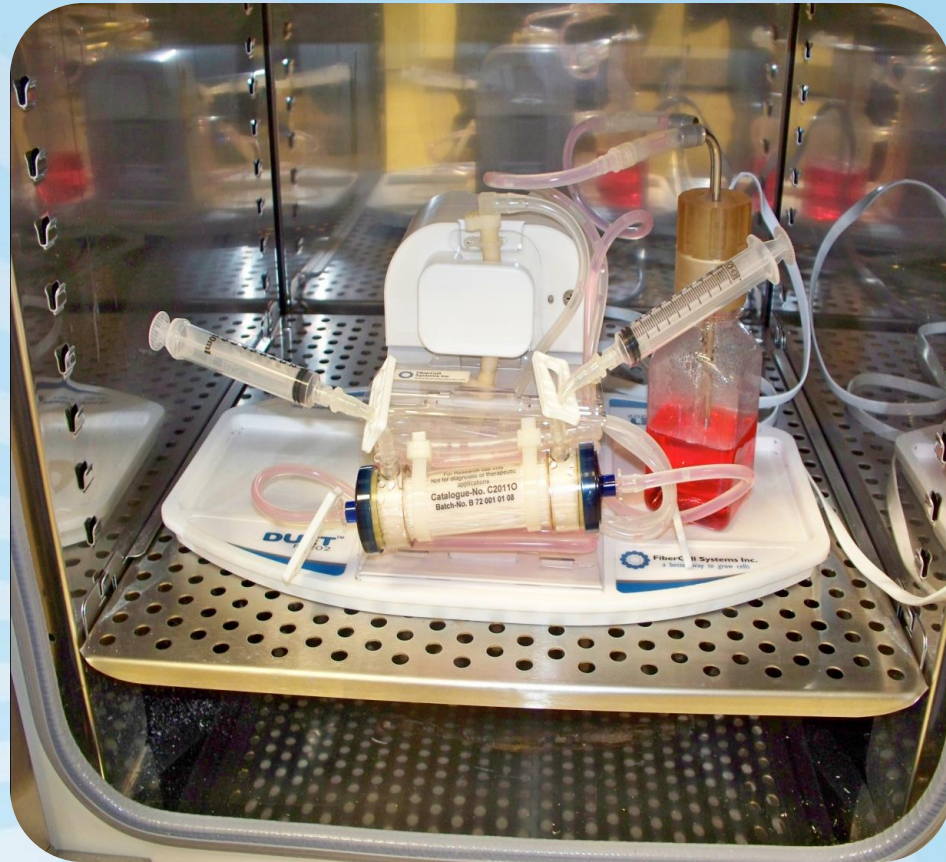


- 10^8 + cells/ml
- “Wavy” fibers optimized for suspension cells
- High cell density permits adaptation to lower serum concentrations and protein free medium such as CDM-HD



- Positive pressure displacement pump
- Silicone tubing for gas exchange
- Closed, bio-safe system

In the Laboratory



- Fits in any standard sized incubator
- Gas controlled by incubator
- Temperature controlled by incubator
- Thin cord for power

Working with the cartridge

- Moves easily into hood
- Good sterile technique always a plus
- Maintenance only 15 minutes per day
- Harvest product and measure glucose consumption

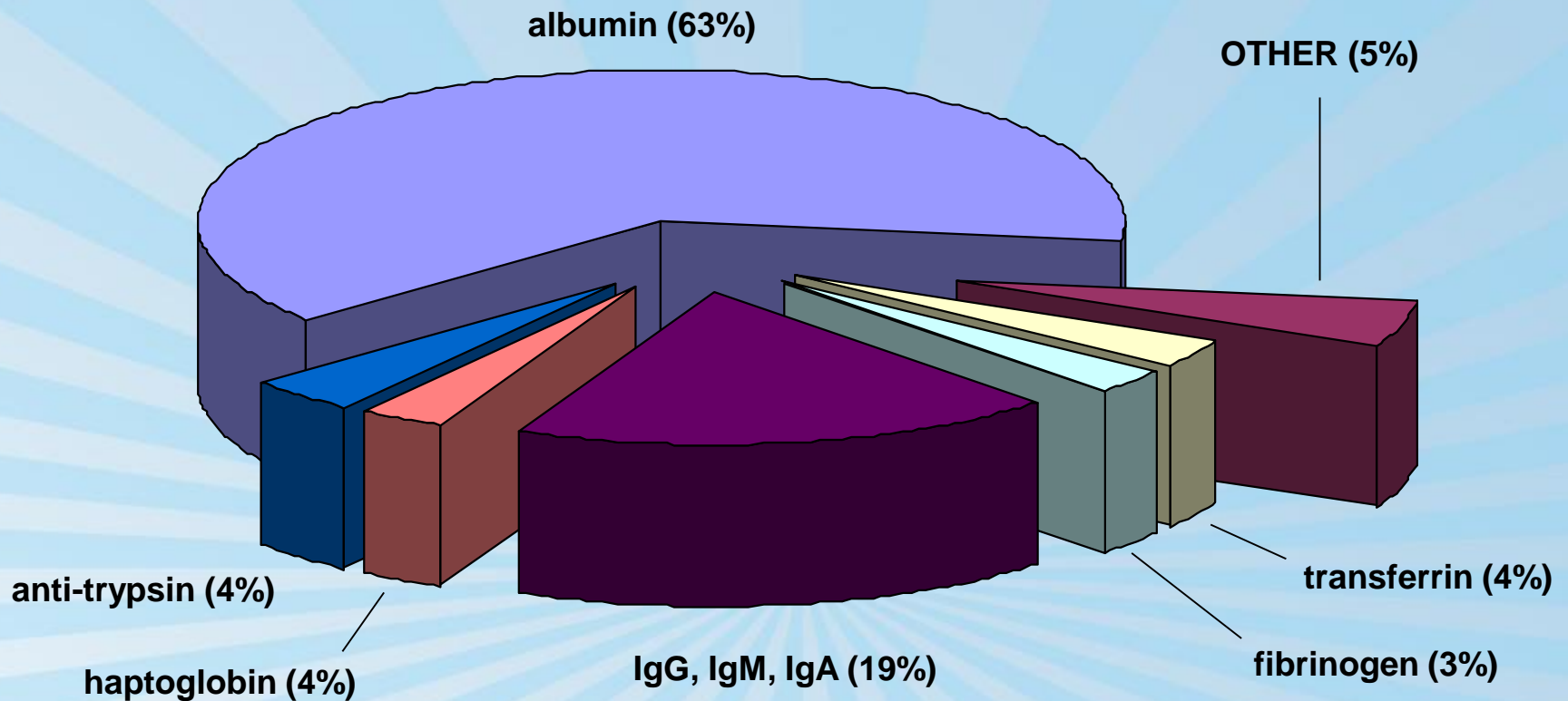


HF Applications

- Monoclonal antibody production
- Recombinant protein production
- Conditioned medium
- Exosome production
- Endothelial cell culture under shear stress
- Cell co-cultivation
- Virus production
- *In Vitro* toxicology

For Research use only!
Not for diagnostic or
therapeutic applications!
Catalogue-No. C2011
Batch-No. B 72 016 06 05

Six proteins constitute 95% of plasma proteins



CDM-HD Serum Replacement



Advantages of Hollow Fiber Cell Culture

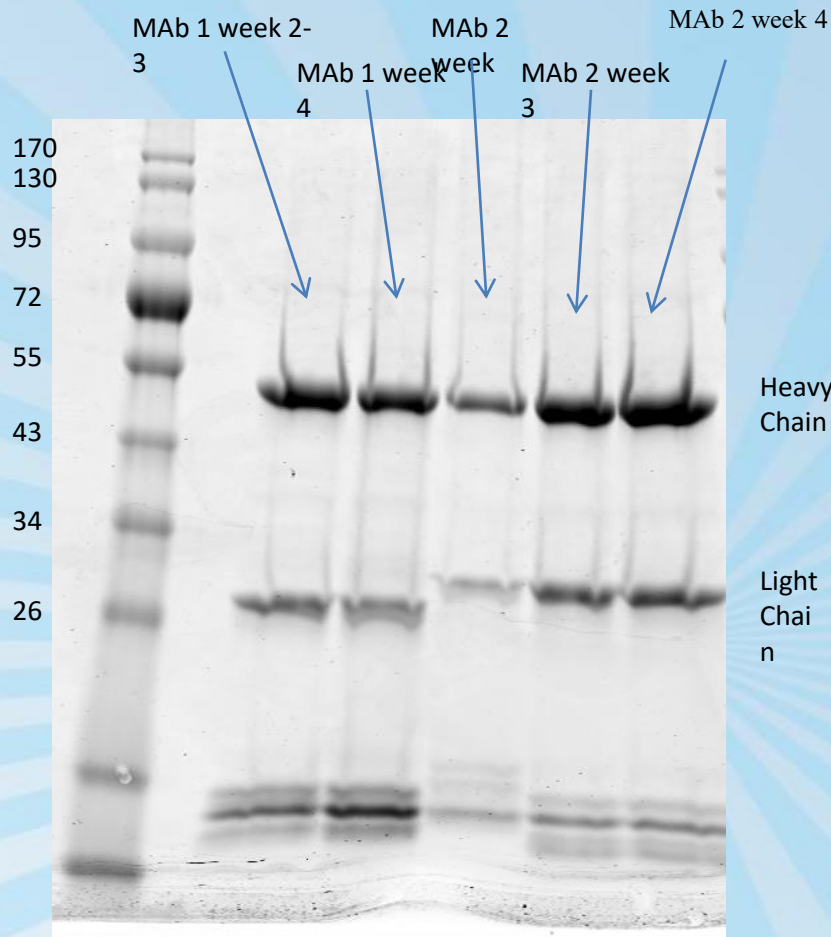
- Concentrated product
- Uniform and complete post-translational modifications
- Low apoptosis, less contamination with intracellular proteins and DNA
- Protein free medium (CDM-HD) contains no surfactants
- Consistency of production over many months.



Monoclonal Antibody Production Using CDM-HD

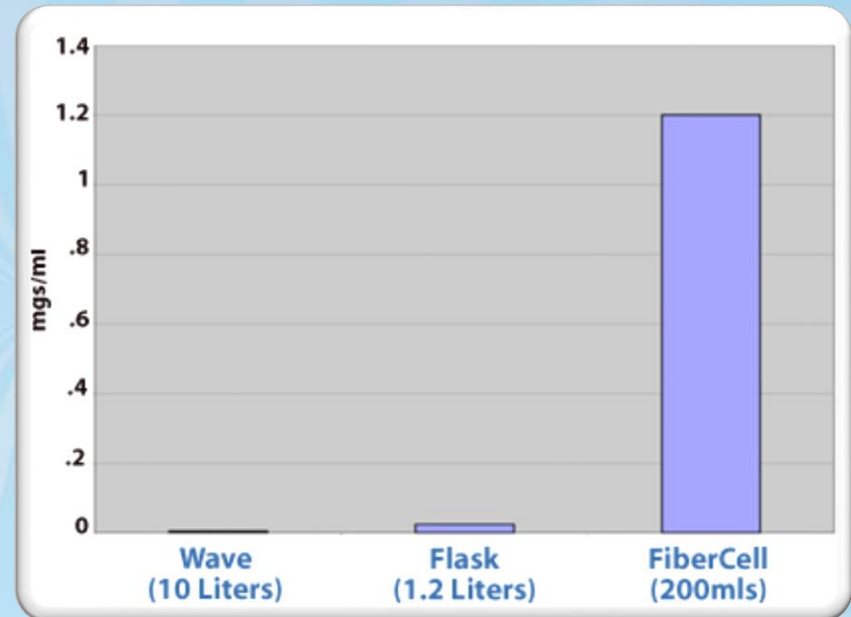
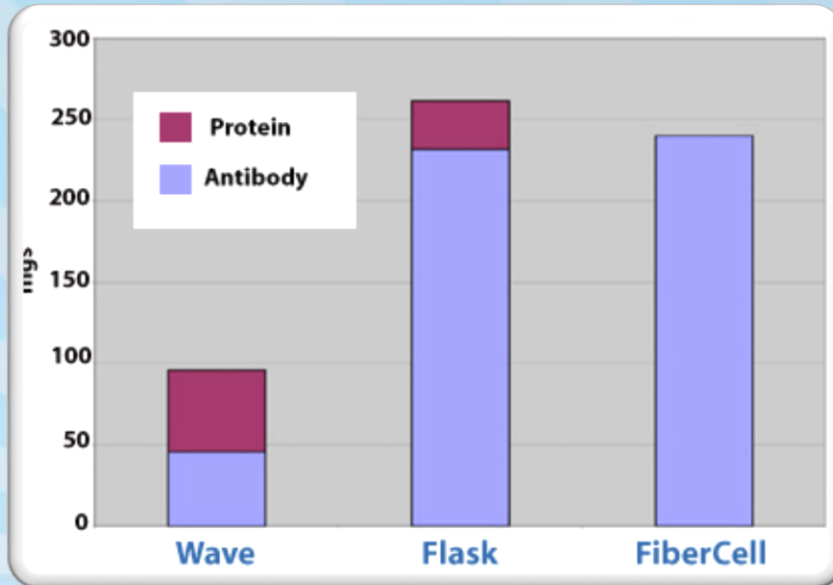
Mab 1: 168 mgs in 60mls volume, 2.8 mgs/ml. 9 liters of medium consumed, three weeks culture

Mab 2: 159 mgs in 70mls volume, 2.3 mg/ml. 11 liters of medium consumed, three weeks culture



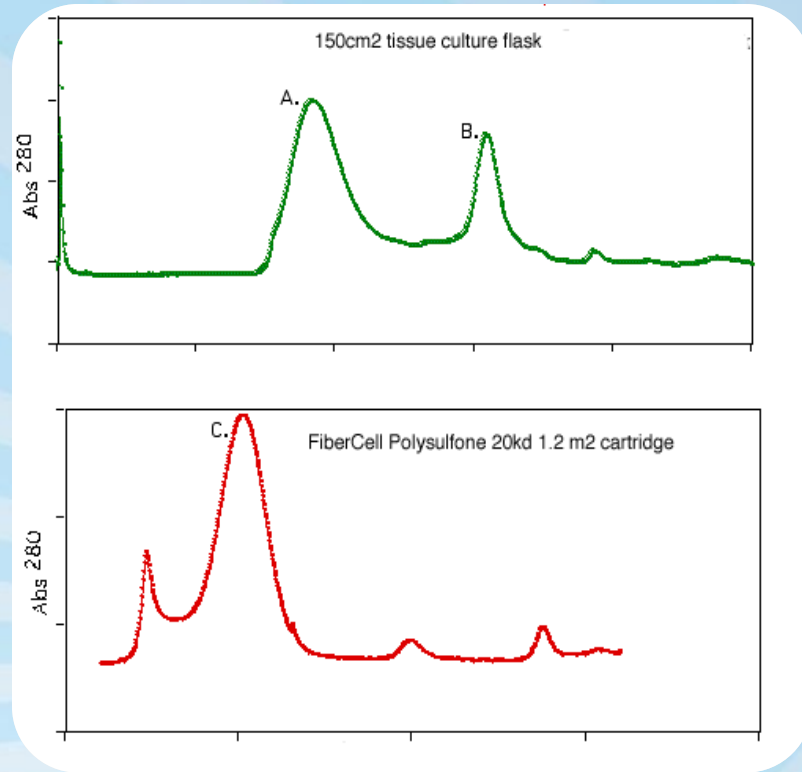
- TGF Beta diffuses out
- MAB trapped in ECS
- Easily adapt to SFM/CDM HD
- Lower endotoxin
- .5 to 5 mg/ml conc.
- 5-100 mg per harvest
- Continuous production for over 6 months

Hollow Fiber/Wave Comparison



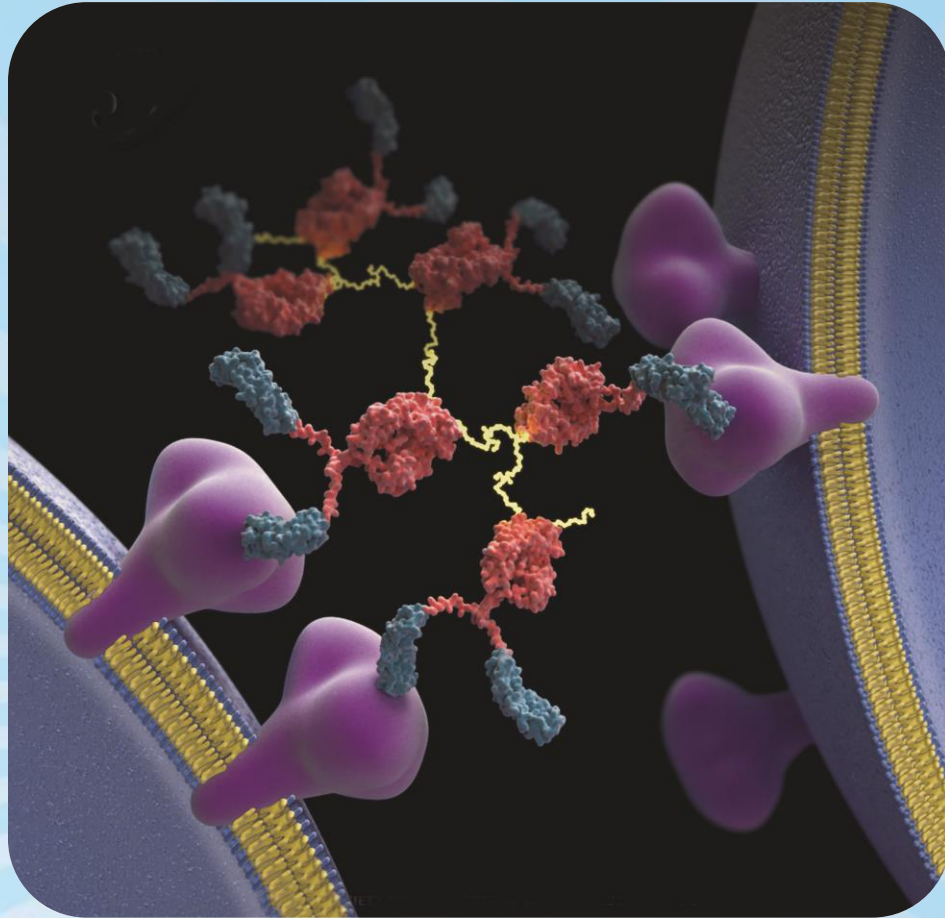
Scale up of hollow fiber is limited by oxygenation

Recombinant Protein Production

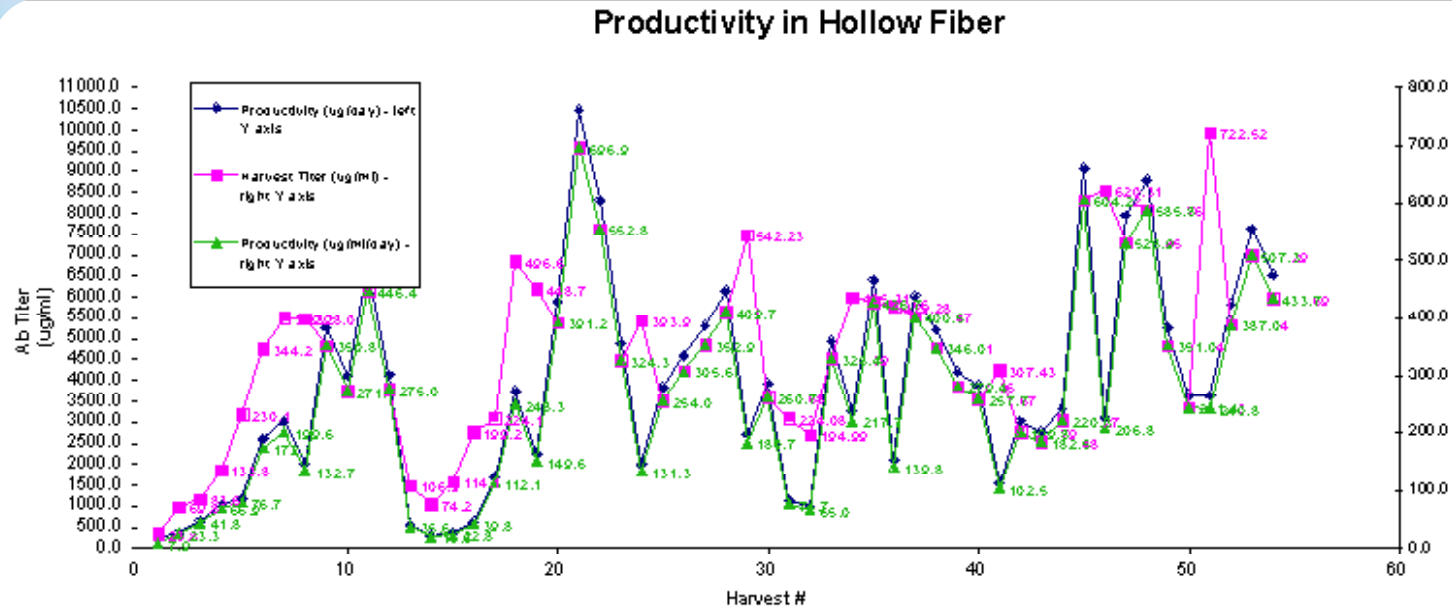


- Both suspension and adherent cell types
- 100X + higher concentration
- Easily adapt to SFM
- Can provide improved protein folding

Journal of Biological Chemistry 9/20/2007

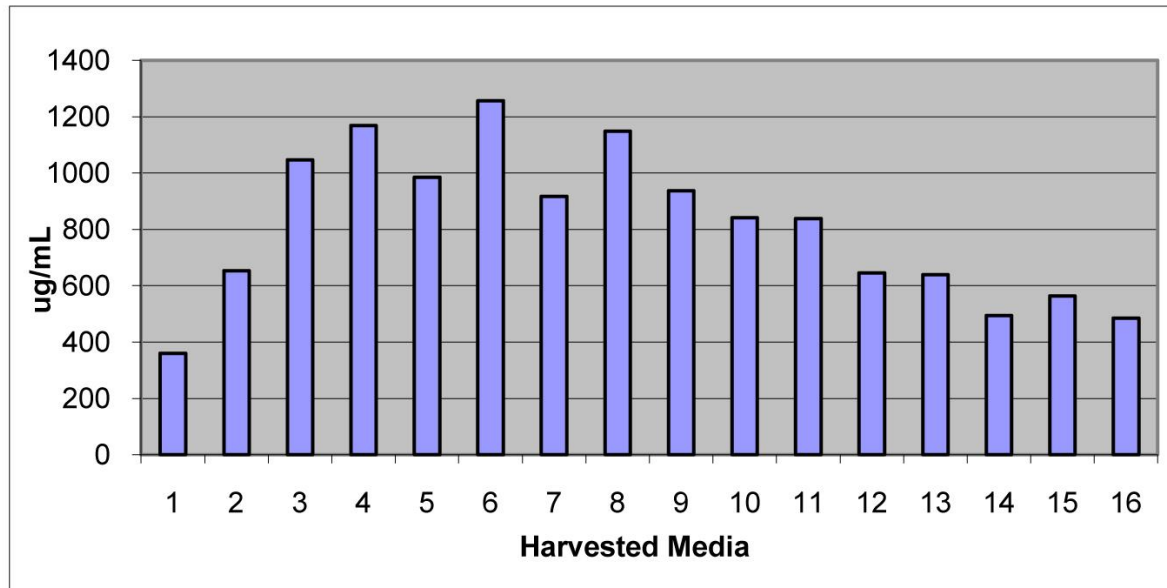


Rec. Protein Production in 293 cells



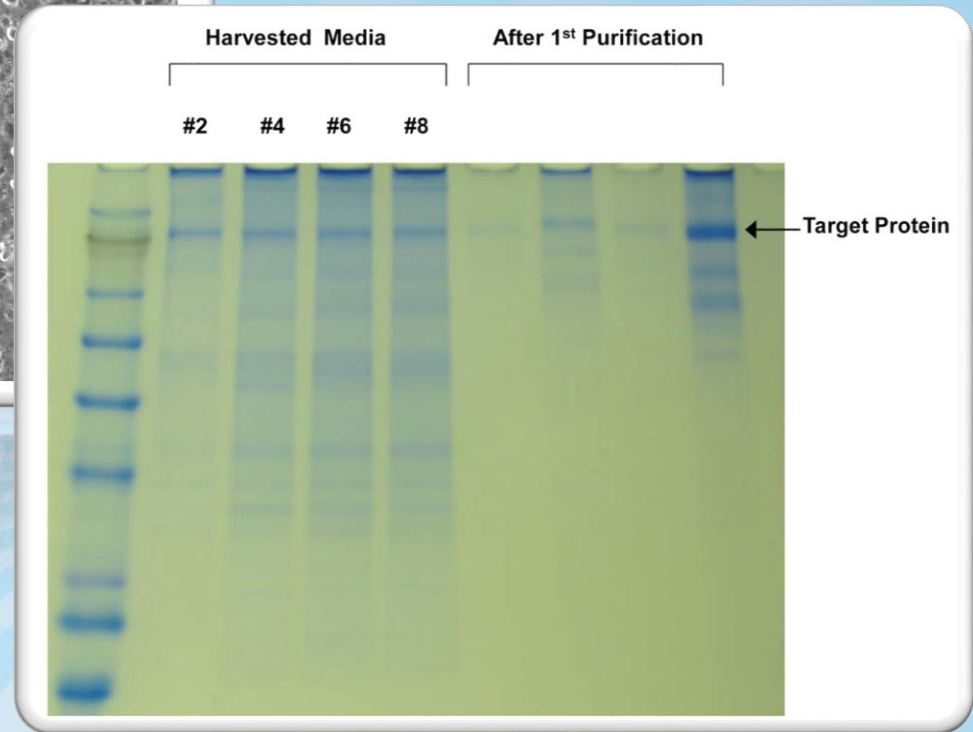
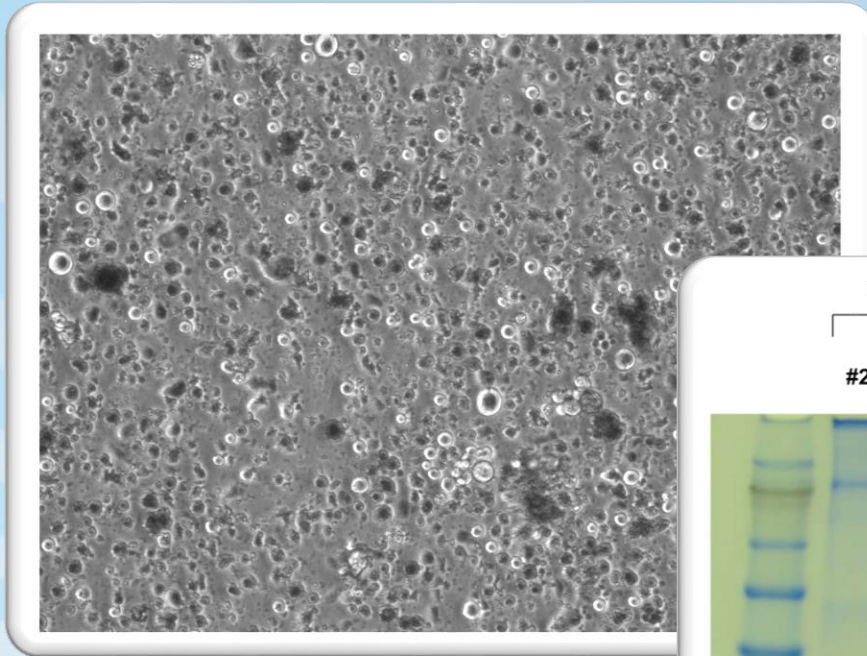
- Rec. IgG from 293 cells
- Produced 276 mg in 2 months using C20 I I
- 900 mL total volume in SFM

Cell Line CHO Suspension Cell (DG44)

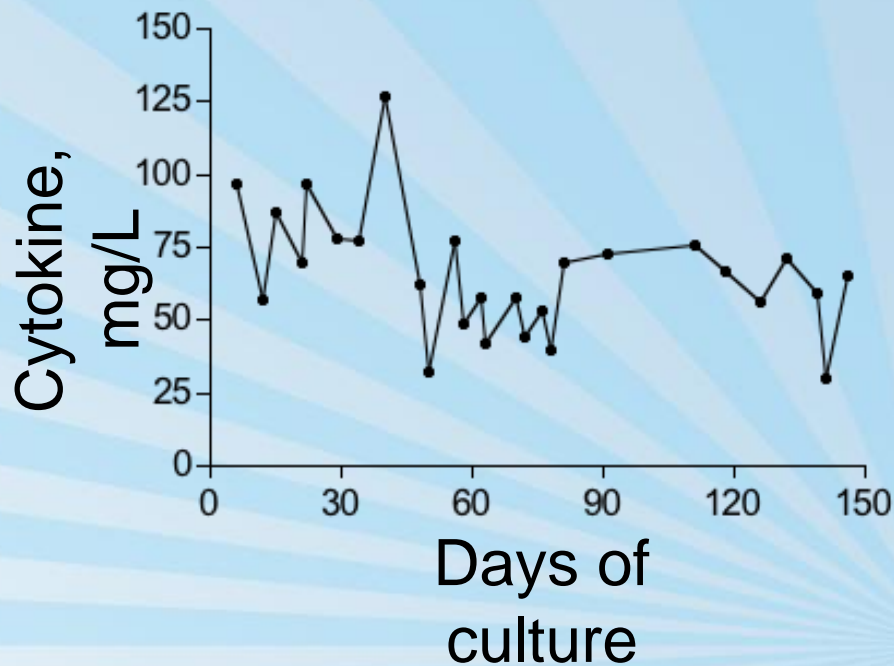


- Protein produced: 246.6 mgs
- Harvest Volume: 304 mls
- Medium consumed: 10 liters
- Culture time: 35 days

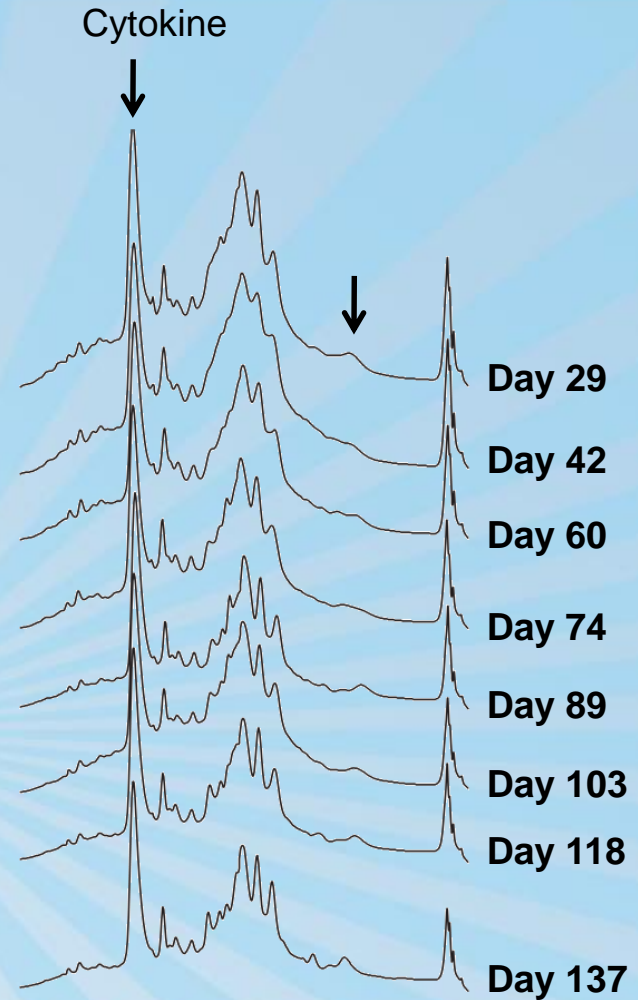
Raw Harvests from DG44 CHO Cell Line



Stable Cytokine Production In Fibercell Hollow Fiber Bioreactor Over 5 Months

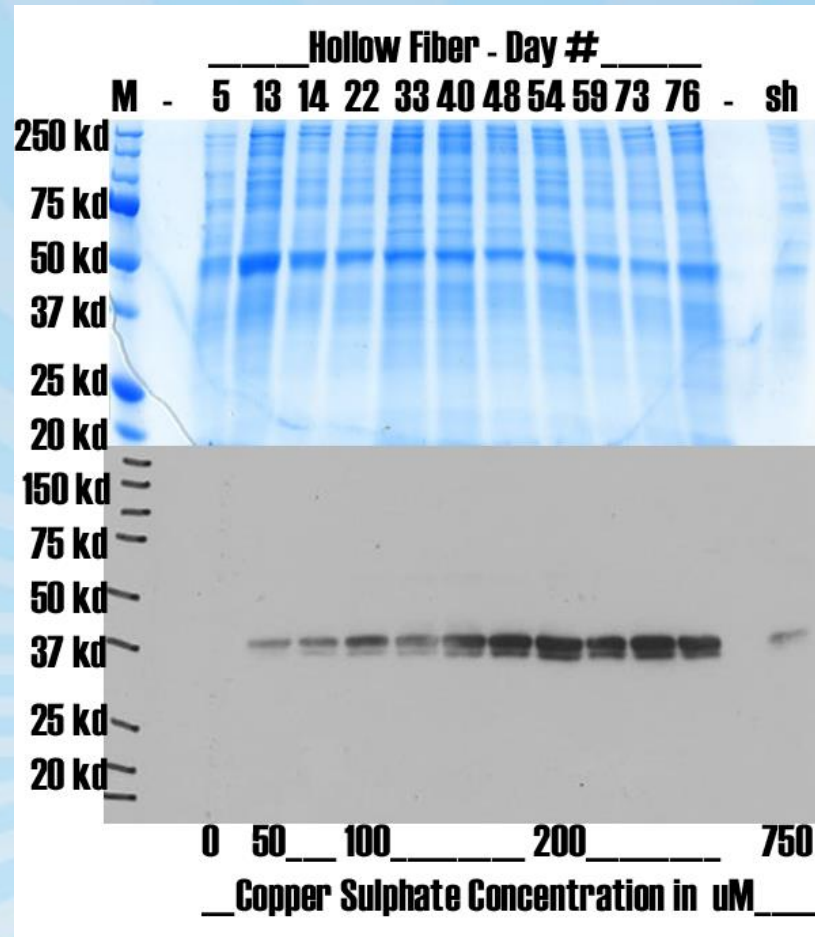


Fibercell Production, mg/Lt



RP-HPLC Analysis, Serum Free Harvest

S2 Protein Production

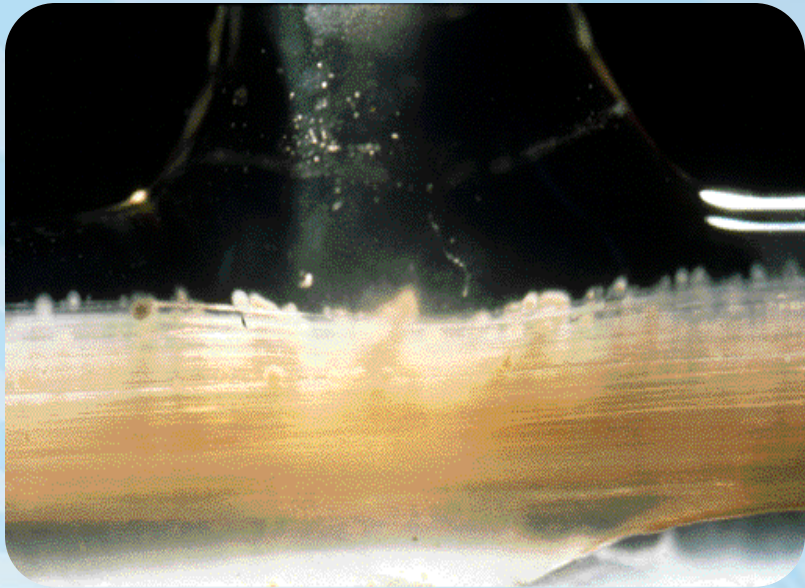


FiberCell Equivalents



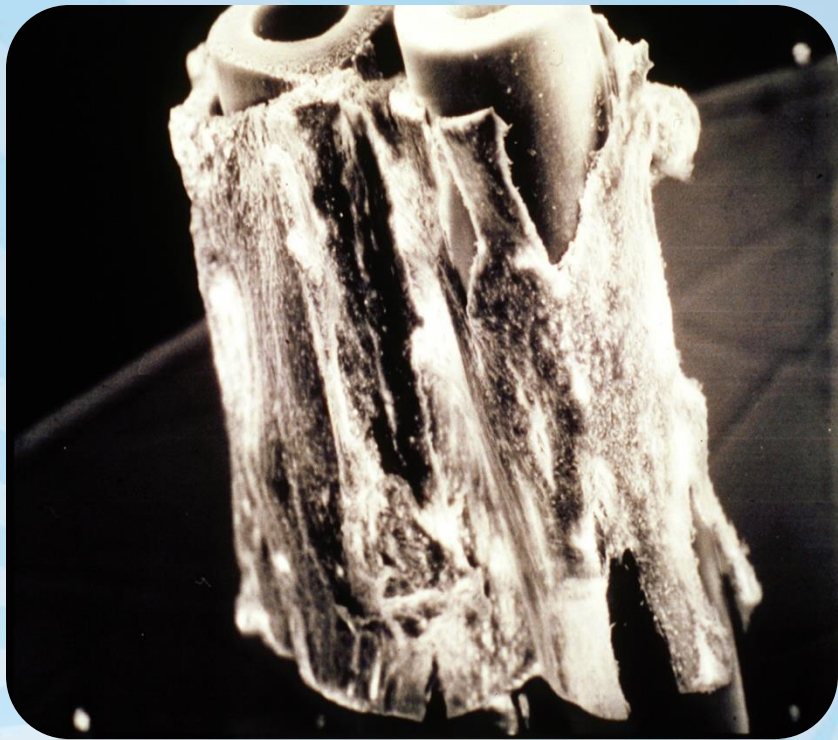
- A single harvest from C2011 can equal 20 roller bottles per day
- A single harvest from C2018 can equal 200 roller bottles per day
- 10mL harvest volume can equal 1 liter of cell culture supernatant

Cellular Co-cultivation



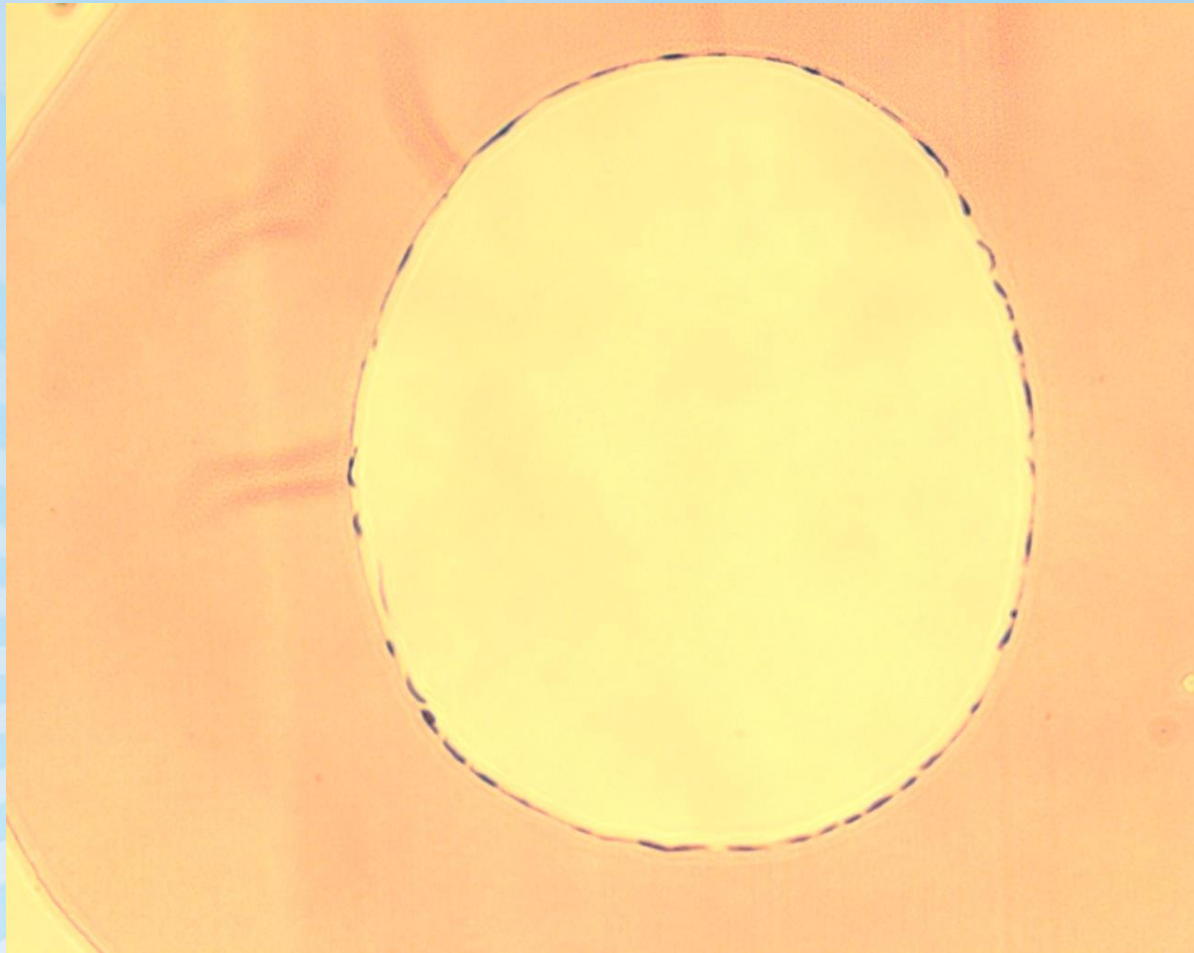
- Only way to get two cell types in close enough proximity and high enough density to observe effects
- Thymic epithelium and thymic fibroblasts co-cultured to form thymic nodules

Asymmetric Cell Co-cultivation

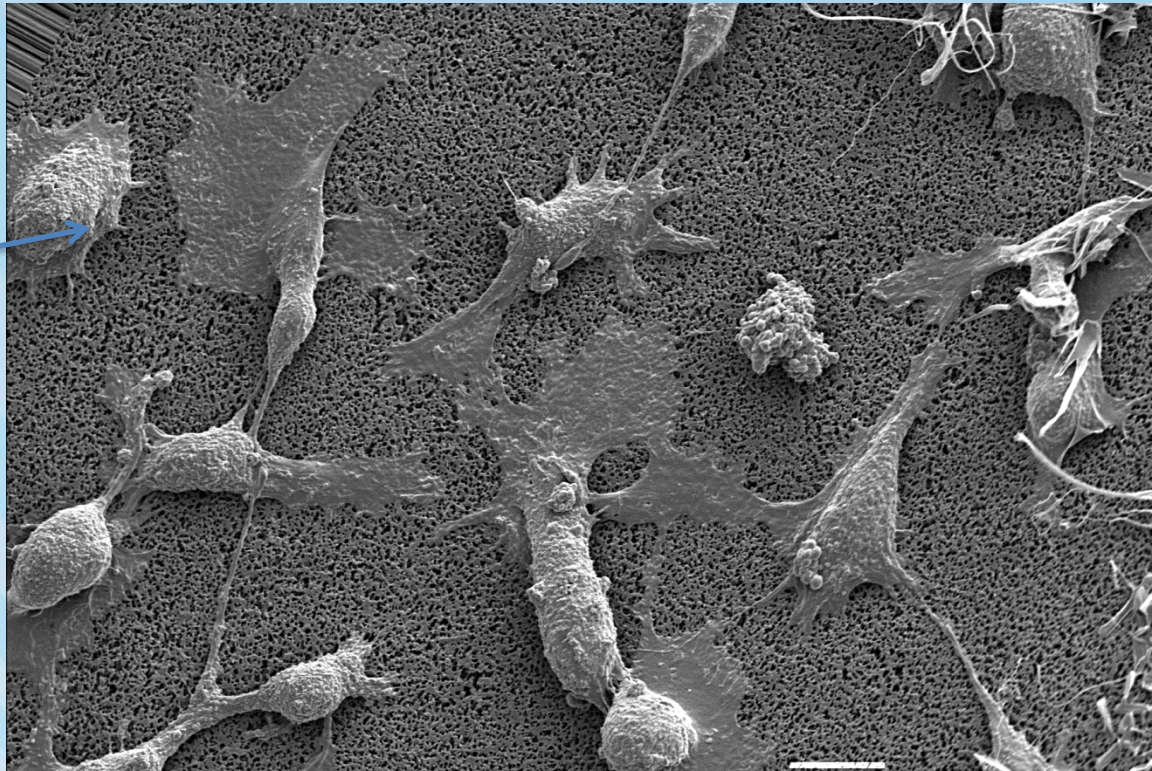


- Co-cultivation of endothelial cells (inside) and vascular smooth muscle (outside)
- Brain endothelial and astroglial cells to form in vitro blood brain barrier

Endothelial Cell Culture

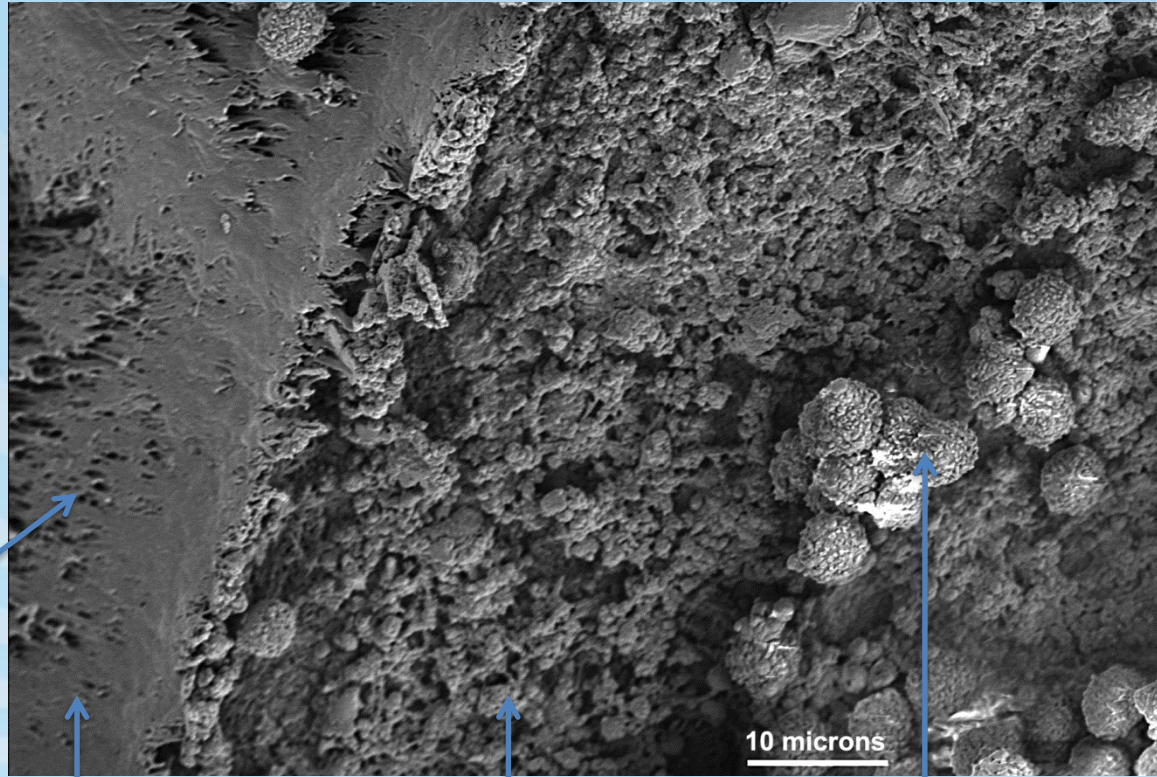


Pores of
fiber



Endothelial cells on the inside on a fiber- these have been adhered to the wall then subjected to very low shear force overnight followed by a few hours at 5 dynes/cm^2 .

While the majority of the cells here are still bulky it is possible to see them begin to flattened down onto the wall of the fiber and really stretch out.



Pores

Fiber

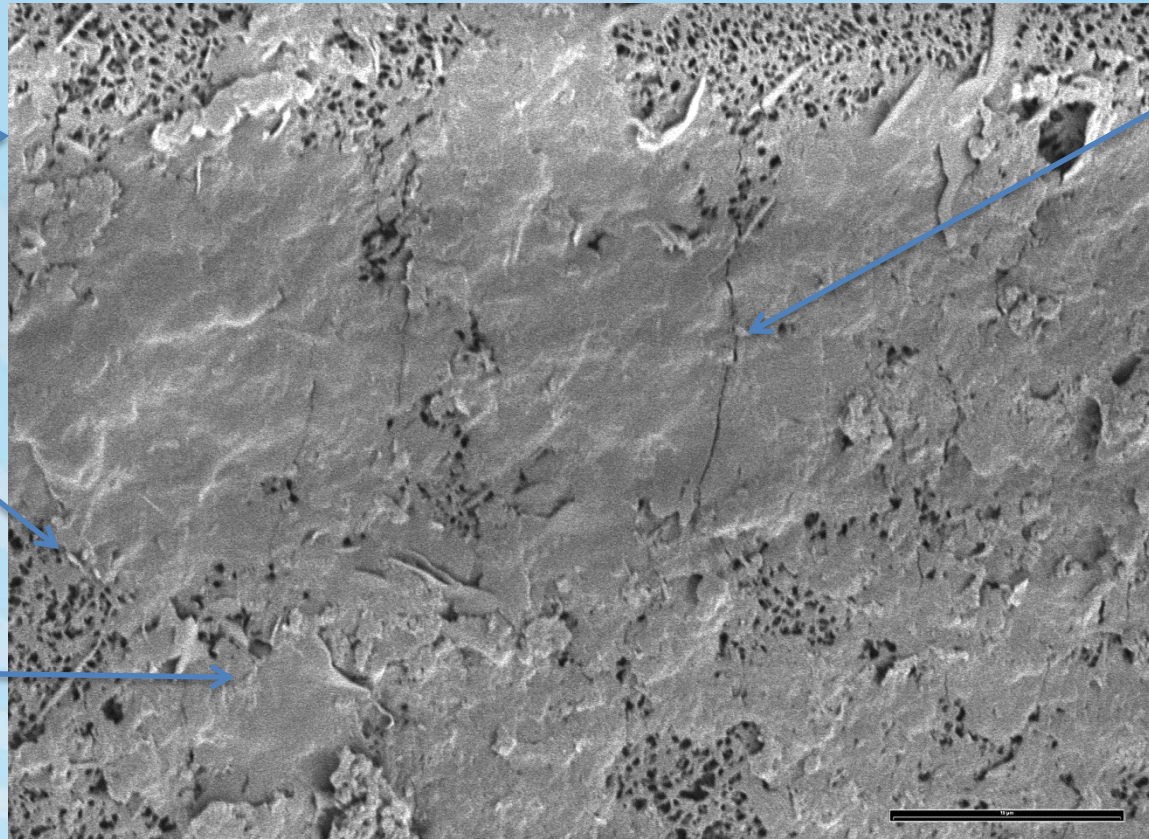
Inside of fiber

CLL cells

Inside of
hollow
fiber

Pore

Endothelial
cell

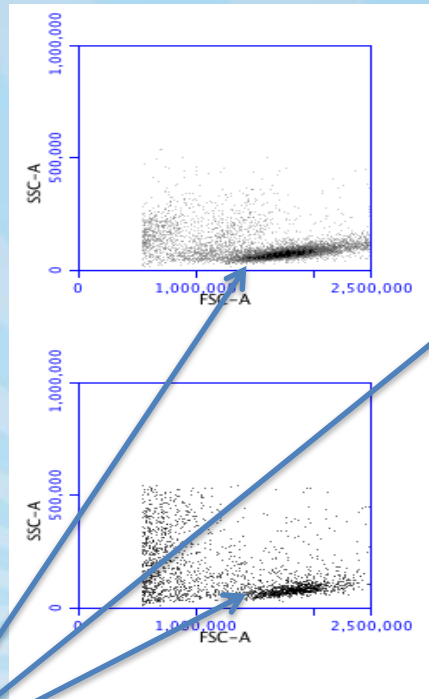


Crack!
Likely to be
a result of
trying to
mount a
sliced
hollow fibre
during
microscopy

Flattened endothelial cells on inside of fiber. These endothelial cells had been adhered to the inside of the fiber and subjected to minimal shear force over night followed by a minimum of 5 hours at 10 dynes/cm². This was based on results we saw when testing the application of a different system that can apply shear force to channels lined with endothelial cells while visible under a microscope. This other system also allowed us to discover that endothelial cells will detach and roll away after alignment if a bubble passes over them so we eliminate all bubbles from the system to preserve the endothelial layer.

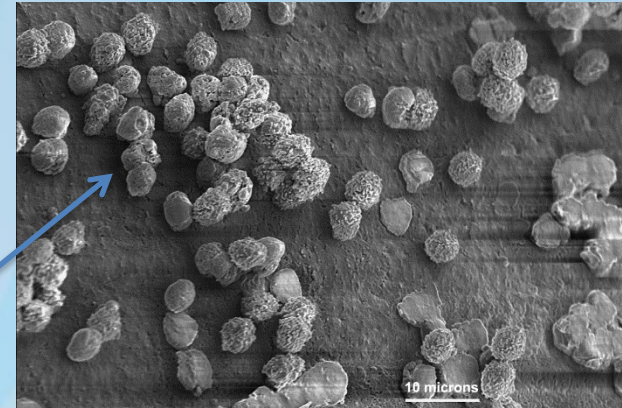
CLL cells actively migrate into the extra-vascular space

Circulating
Compartment



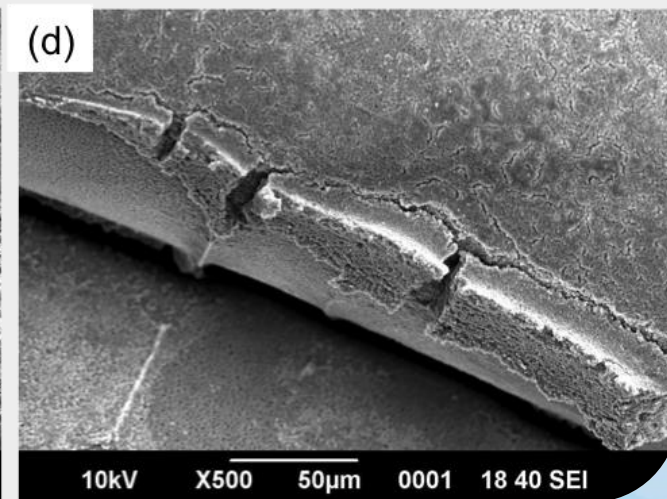
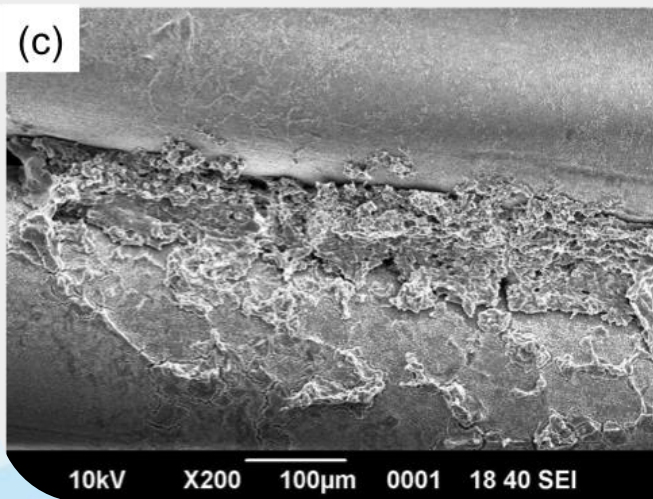
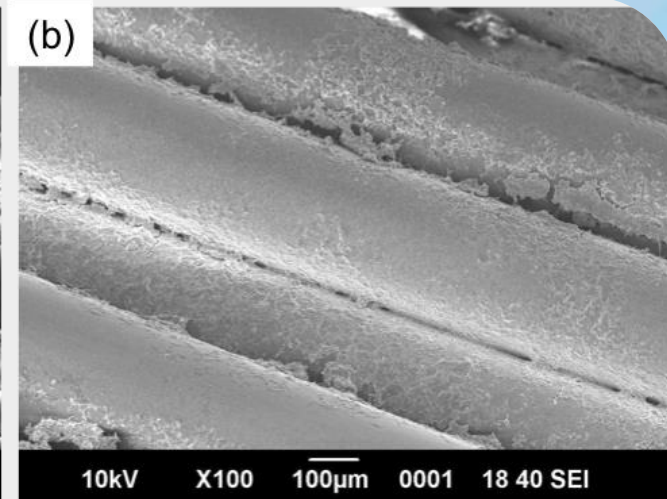
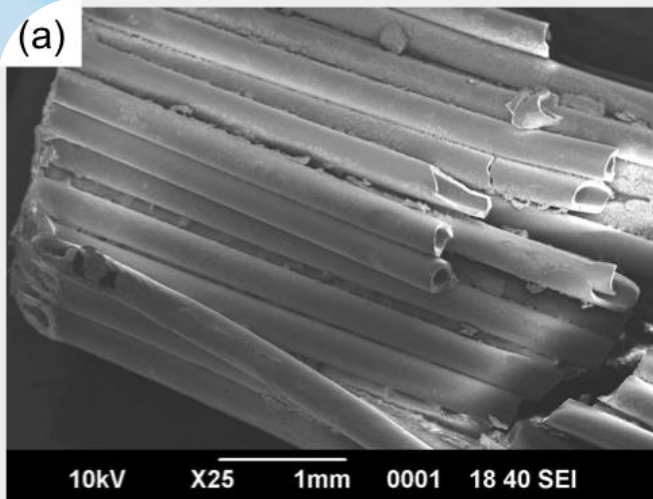
Extra-vascular space
(migrated
compartment)

CLL
cells



Scanning electron
micrograph of the
outside of a hollow
fibre after circulation
of CLL cells around
the system

Bone Marrow Stroma/HSC Co-Culture

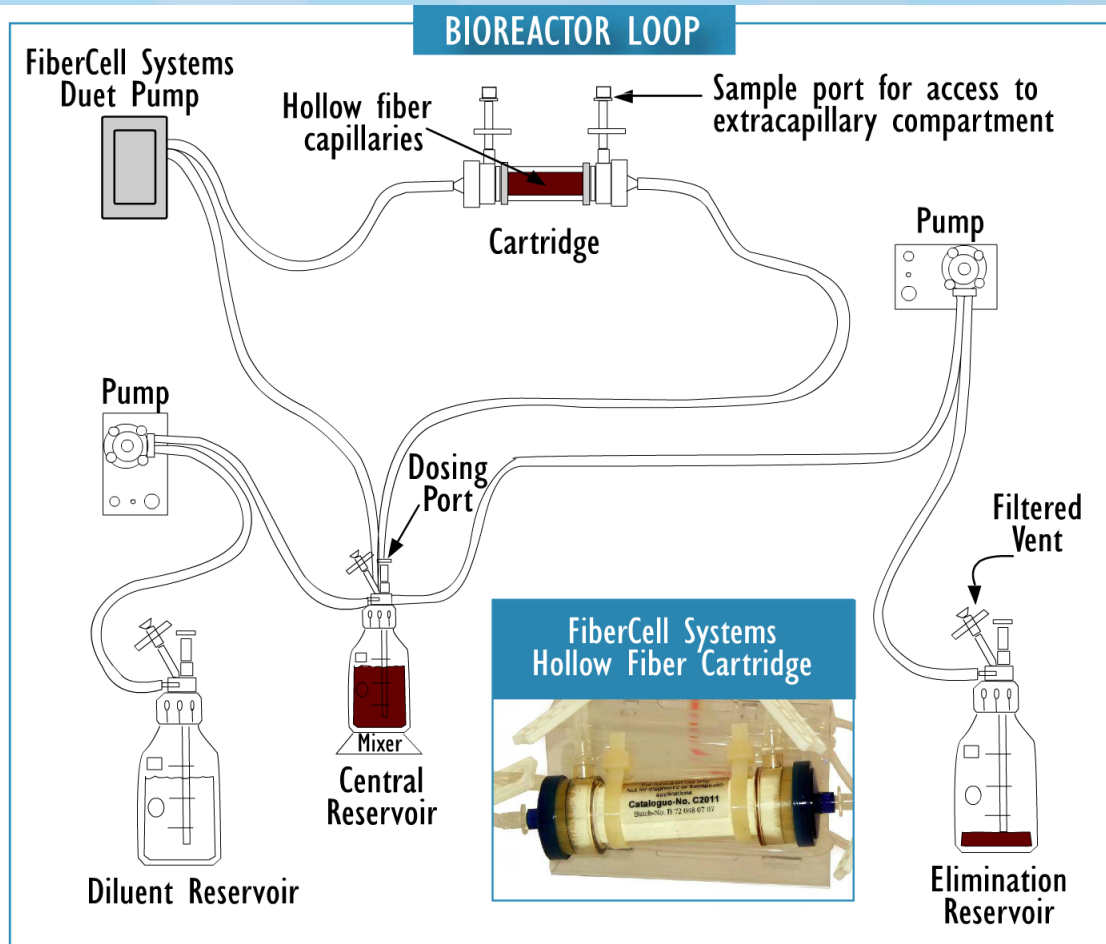


Lack of new antibiotics



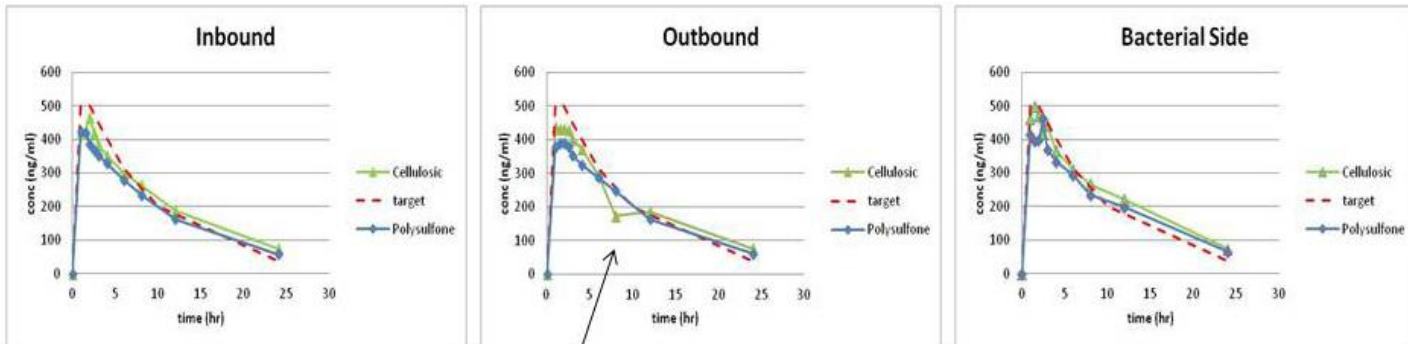
- Only 2 systemic antibiotic agents approved since 2008
- 16 approved between 1983 and 1987
- 3 reasons:
 - **Scientific:** Easy to discover antibiotics have already been found
 - **Economic:** Antibiotics represent a poor return on investment and new antibiotics reserved for difficult cases
 - **Regulatory:** FDA approval process increasingly complex and expensive.

Hollow Fiber Infection Model

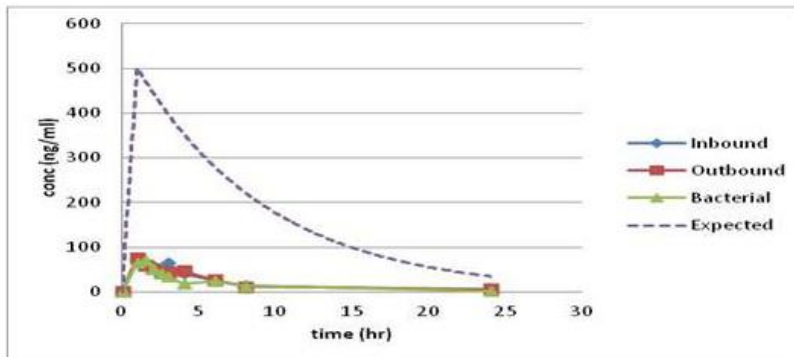


PK Profile

This drug showed good compatibility with both cartridges.



Not significant enough to consider incompatibility



This drug is not a good candidate for HFS use

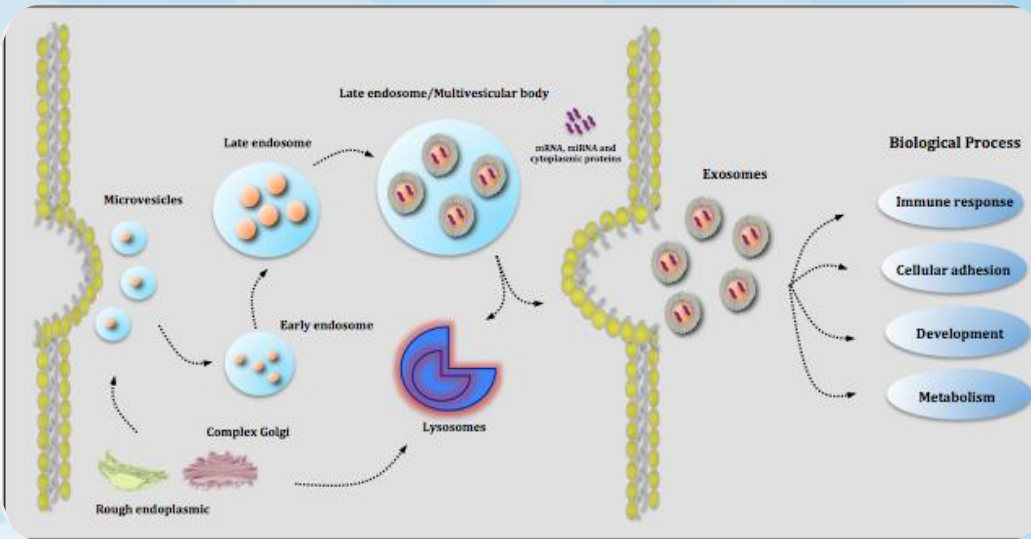


The hollow fiber infection model is a complementary and additional tool for drug development, to be implemented at the earliest stages

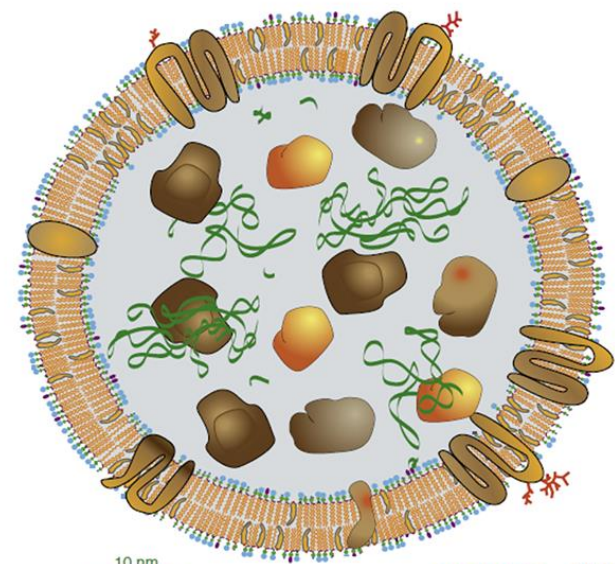
- Optimal dose selection and route of administration
- Optimal dosing schedule
- Possible combination therapies
- Defines emerging resistance
- Defines total kill
- Post-approval drug regimen optimization
- Can support trial design for Phase I, II, III and IV clinical trials

Exosomes

- Cell-derived vesicles in biological fluids
- Including medium of cultured cells
- Dia. between 30 and 100 nm
- Contain cellular proteins and RNA
- Facilitate cell-to-cell transfer of cargo
- May play a role in cell-to-cell signaling
- May mediate adaptive immune responses



The Anatomy of an Exosome



- Ceramide
- Sphingomyelin
- Phosphatidylserine
- Phosphatidylethanolamine
- lyso-Phosphatidylcholine
- Phosphatidylcholine
- Phosphatidylinositol
- Cholesterol

Human Adipose Derived Adult MSC

130 T225 Flasks

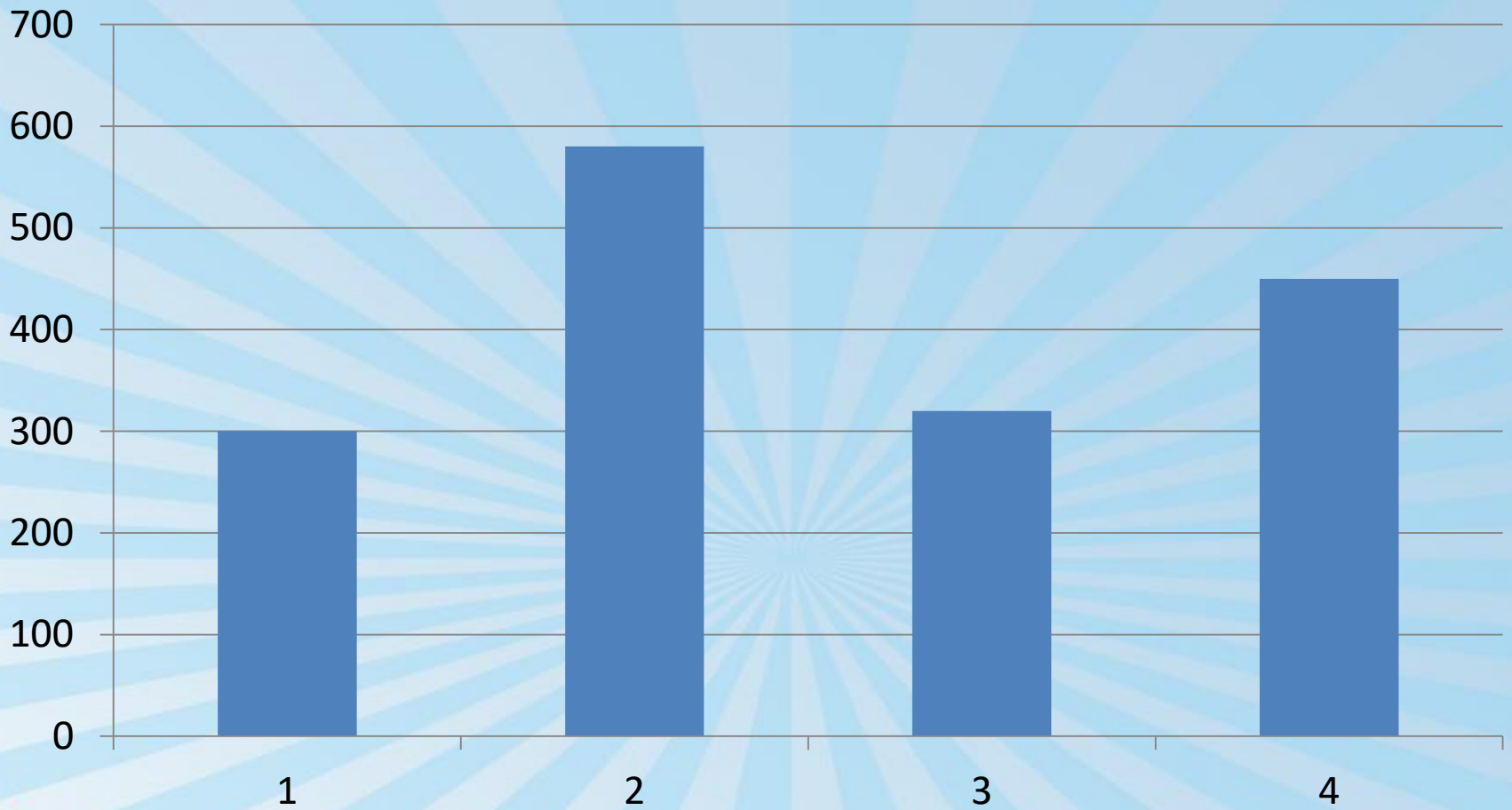
- Harvest Volume: 4000mls
- Protein: 0.9 mgs
- Particles: 1.6×10^9

One C2011 Bioreactor

- Harvest Volume: 120mls
- Protein: 14.45 mgs
- Particles: 3.27×10^{12}

5×10^8 cells seeded

Culture Platform (1 x 10⁹ cells)	Medium Volume (mL)	Total Number of EVs (10⁹)
Bioreactor Harvest #1	40	320
Bioreactor Harvest #2	40	250
Bioreactor Harvest #3	40	290
130 Flasks (225 cm ²)	4000	16





Advantages for Exosome Production

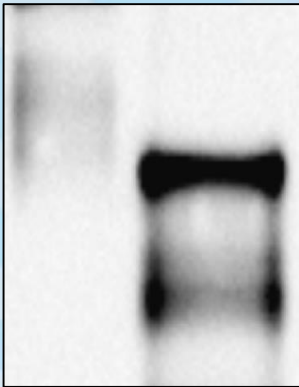
- Large numbers of cells can be cultured in a small space
- Secreted exosomes are concentrated
- Continuous production over several months
- Serum can be used without contamination from endogenous exosomes
- CDM HD can be used for cGMP production
- Cell proliferation may be limited



Bioreactor is a rich source of exosomes

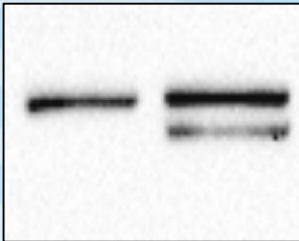
Flask
exosomes

Bioreactor
exosomes



CD63

**7.6-fold
increase**

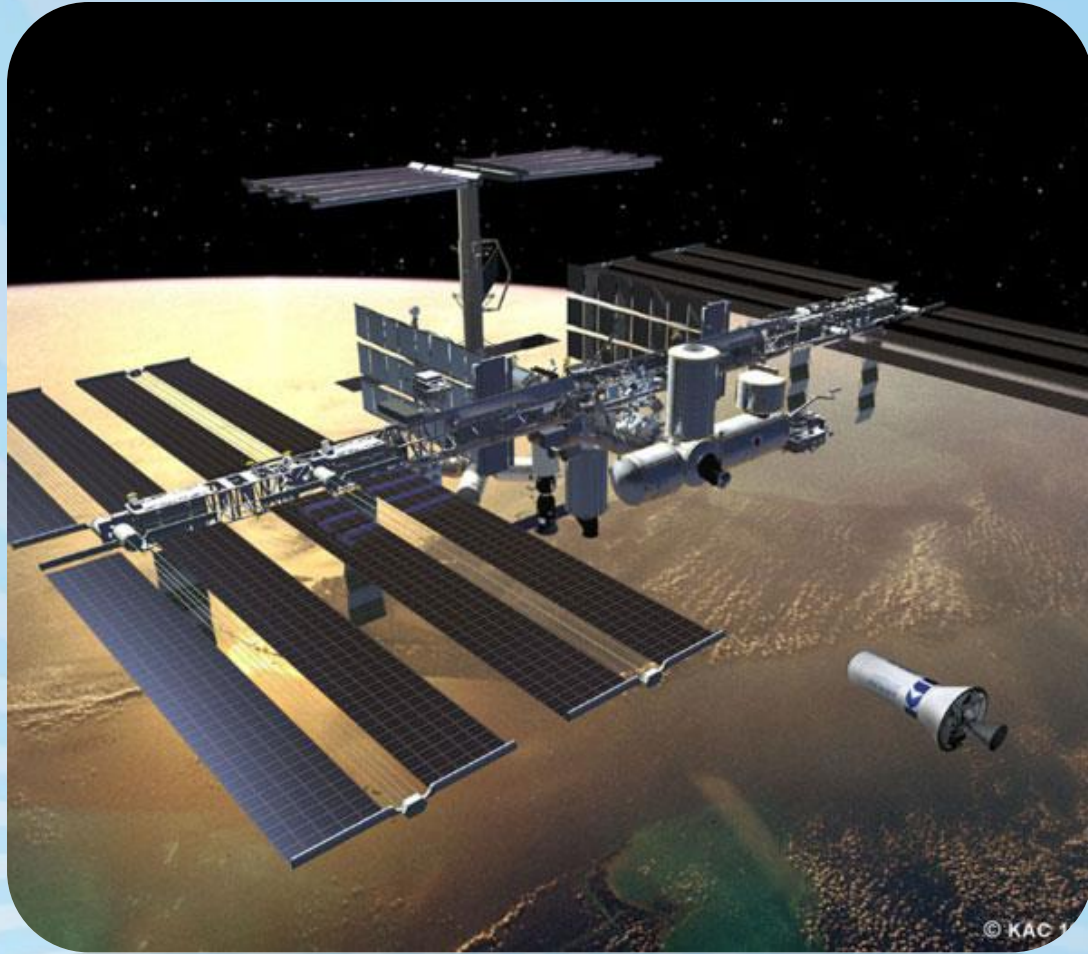


Alix

**2.1-fold
increase**

Sample	μg yield per ml sup
Conventional culture EVs (comparison protocol)	2.7 ± 0.4
Bioreactor EVs (comparison protocol)	33 ± 3
Bioreactor EVs (optimized protocol)	54 ± 2

**>10-fold increase in exosome yield
from bioreactor supernatants**



© KAC 1

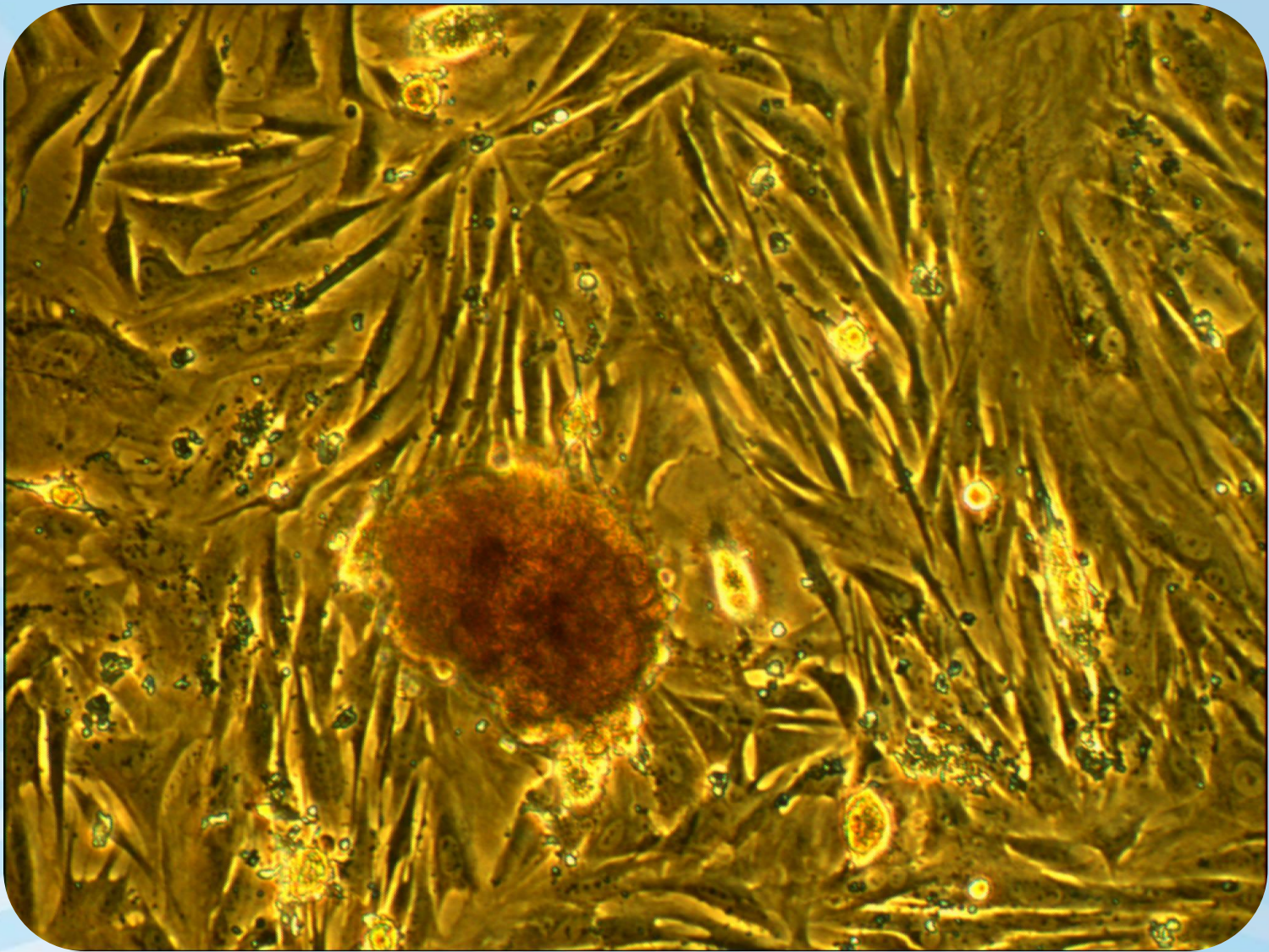
© KVC 1

Pulsatile Perfusion of Placenta



Placental Co-Culture





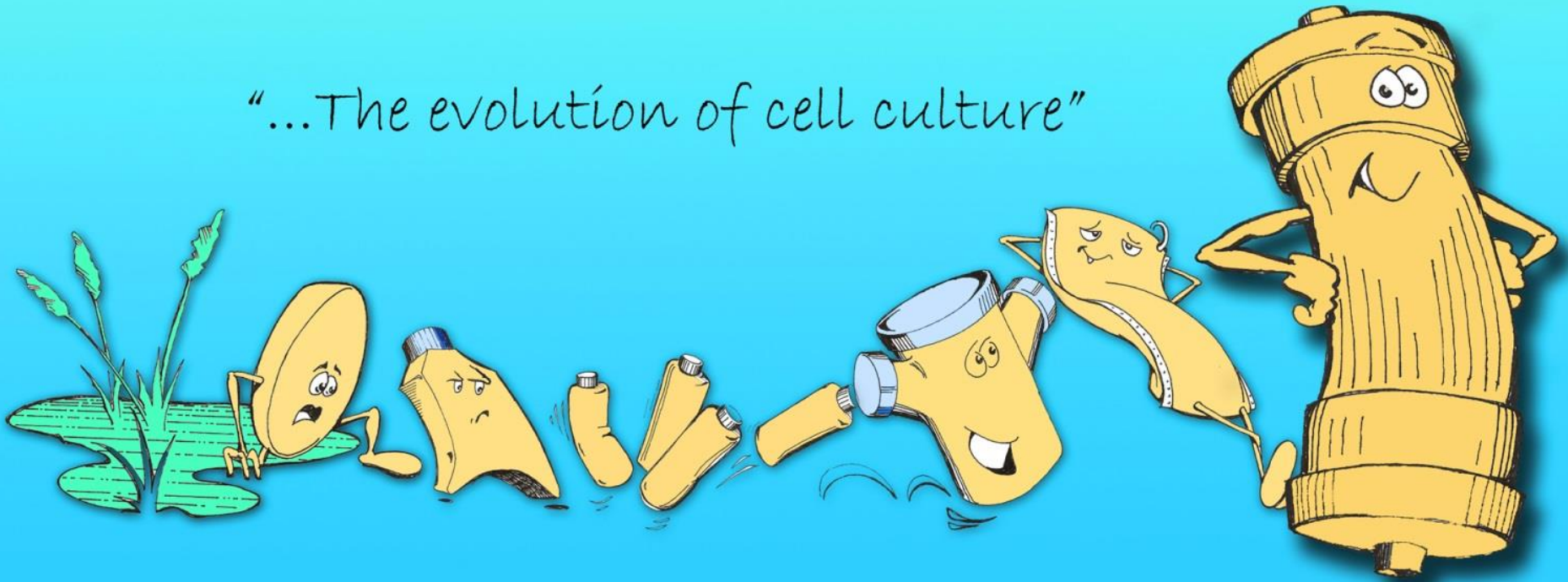
Harvest vs. Flask

Phenotype	ECS Harvest	Flask
• CD 45	4%	1%
• CD 34	0%	0%
• CD133/2	2%	0%
• CD31	3%	48%
• CD 13	6%	83%
• CD 105	43%	99%
• CD 73	18%	99%
• CD 90	5%	96%
• CD 14	23%	4%
• NANOG	0%	0%

Summary

- Hollow fiber bioreactors are the method of choice for the culture of 10^9 to 10^{11} cells
- Ideal for producing 100mgs to several grams of MAB, 10mgs to 100s of mgs of recombinant proteins
- Concentration of products 10 to 100X higher than with conventional methods
- The most *in vivo* method for culturing cells over long periods of time
- Saves time, space, purification costs

“...The evolution of cell culture”



Thank you.







Recombinant Protein and Monoclonal Antibiotic Production in Hollow Fiber Bioreactors

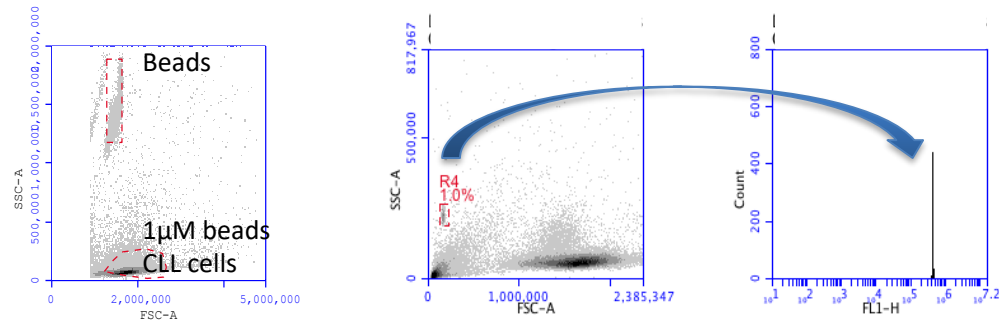
Lab Scale To Process Scale

By John J.S. Cadwell

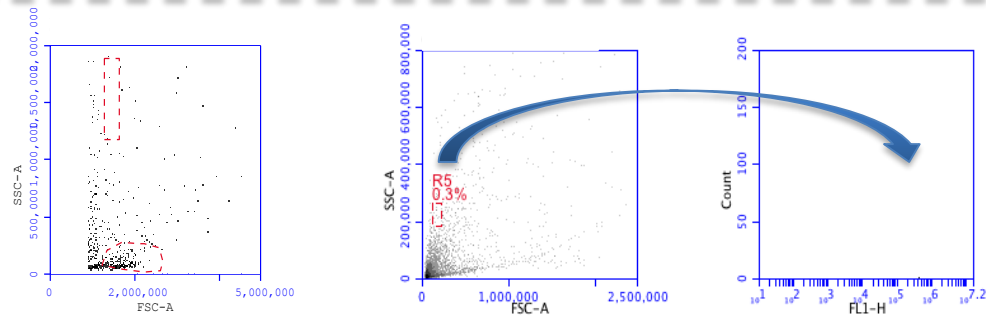


CLL cells actively migrate into the extra-vascular space

Circulating
Compartment



Extra-vascular
space (migrated
compartment)



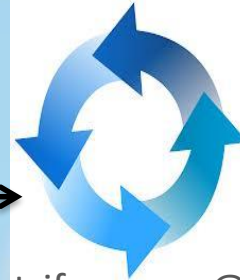
Exosome isolation via ultracentrifugation



60ml (3 ea.)
FiberCell harvests,
cleared of cells



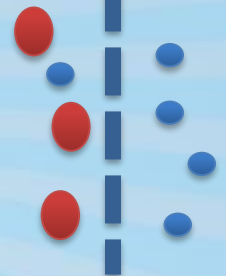
Centrifuge sup @:
3,000xg (15min) to
remove cells and
debris



Centrifuge sup @:
20,000xg (30min) to
remove large vesicles



Filter through
0.22 μ m



100kDa MWCO for
~10x concentration

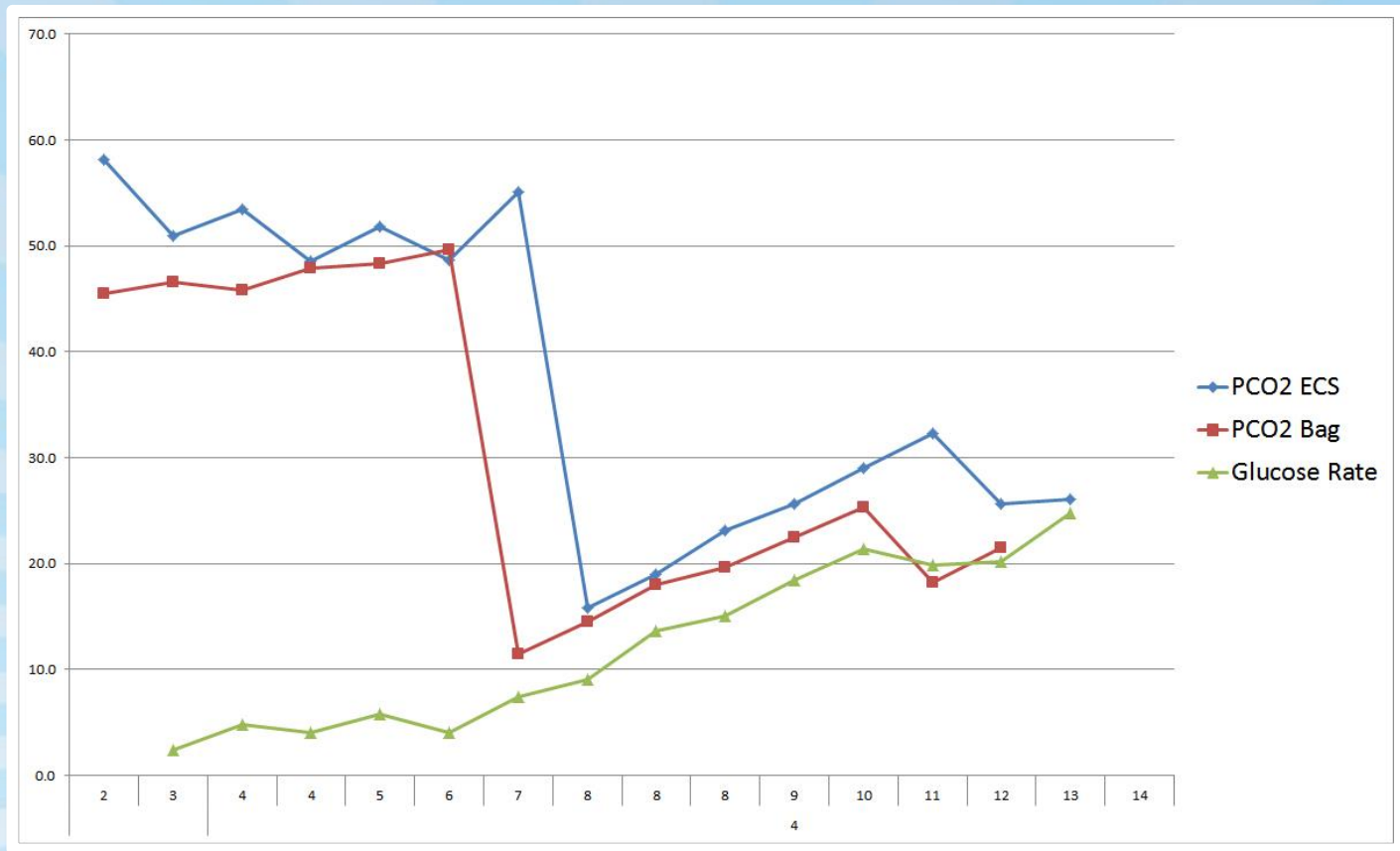


- Pellet exosomes by 110,000xg ultracentrifugation for 2h
- Wash by resuspending in 6ml PBS, and repelleting @120,000xg for 90min

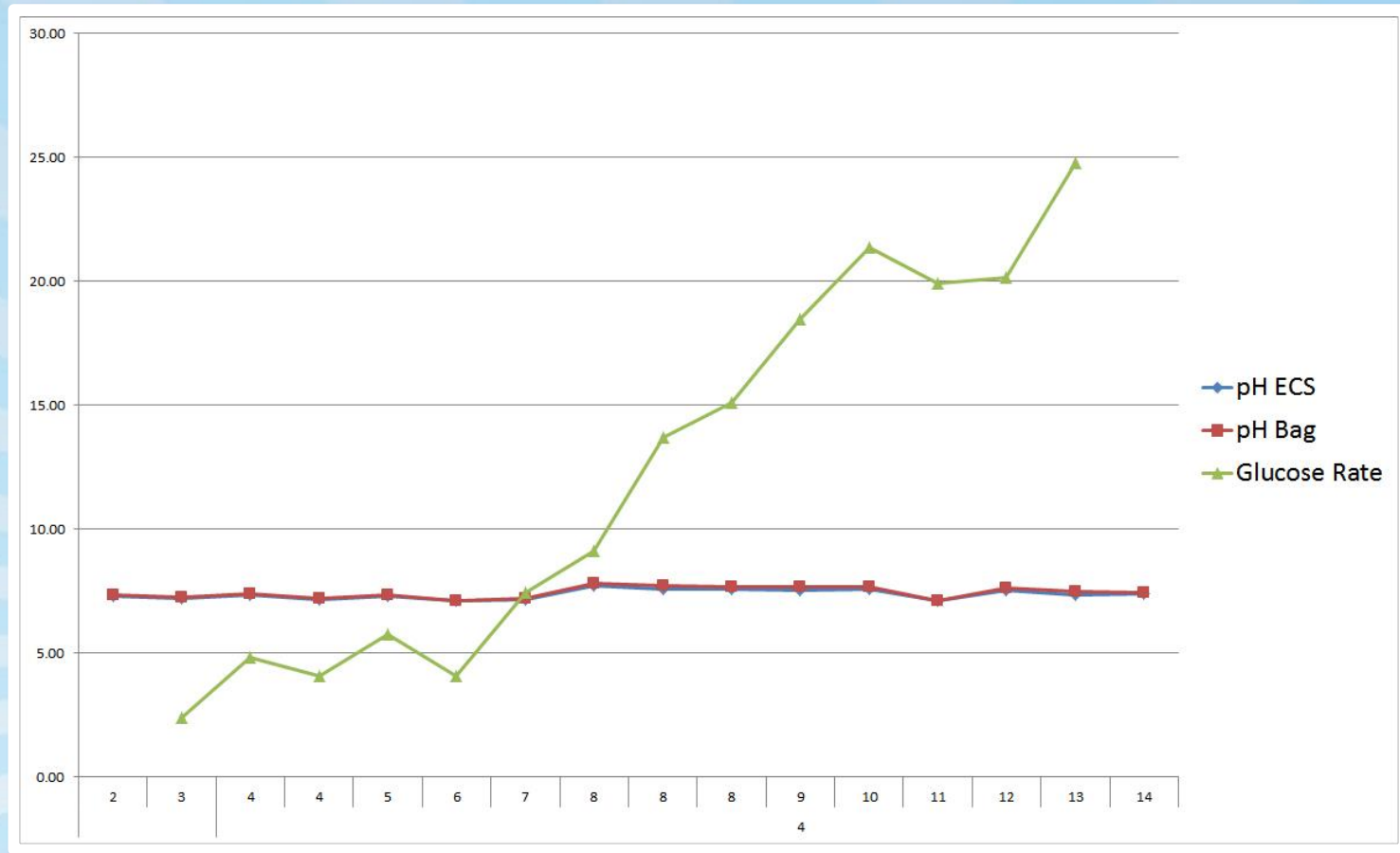
Resuspend in
1ml sucrose
buffer*



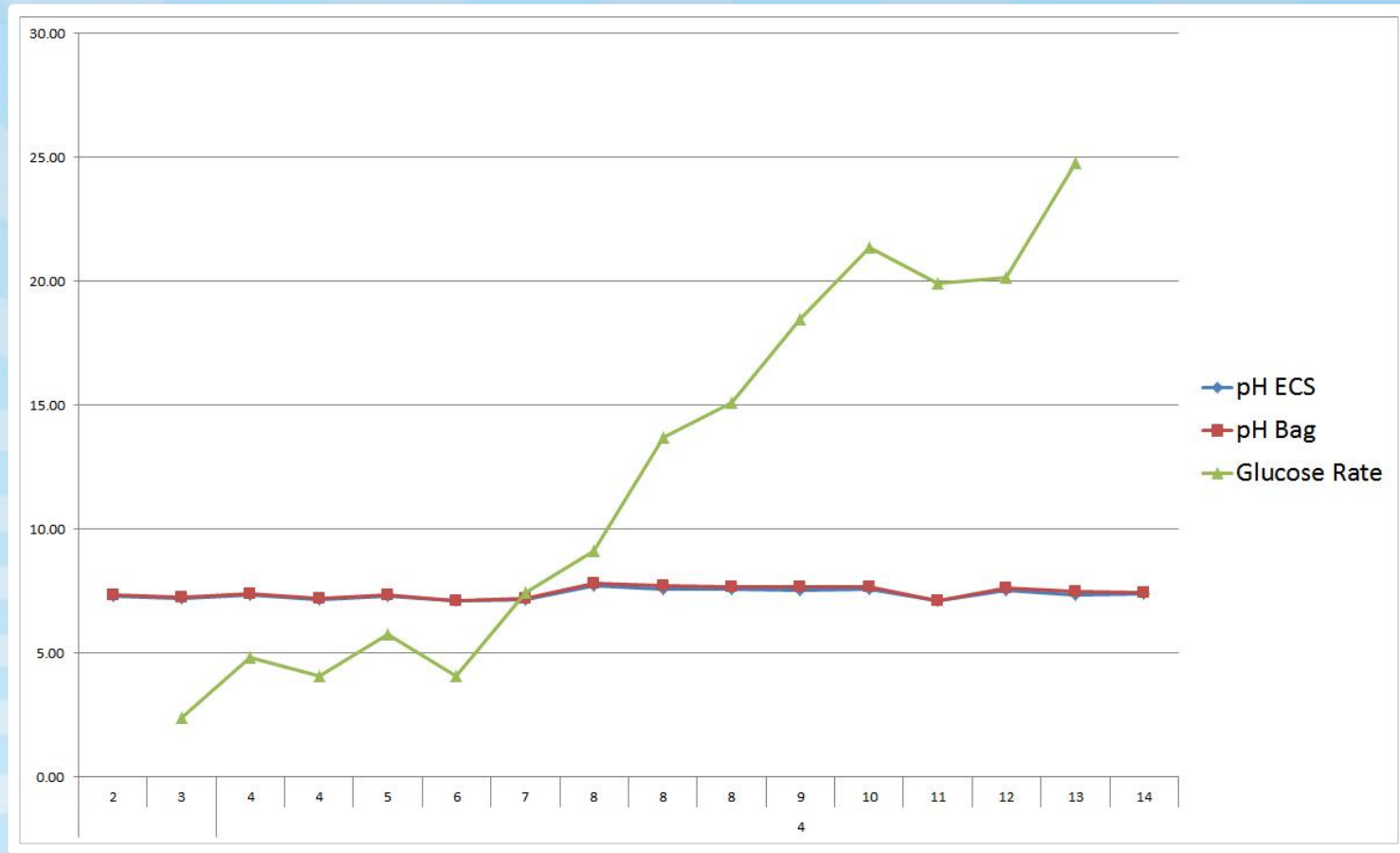
Data showing exchange across the fiber



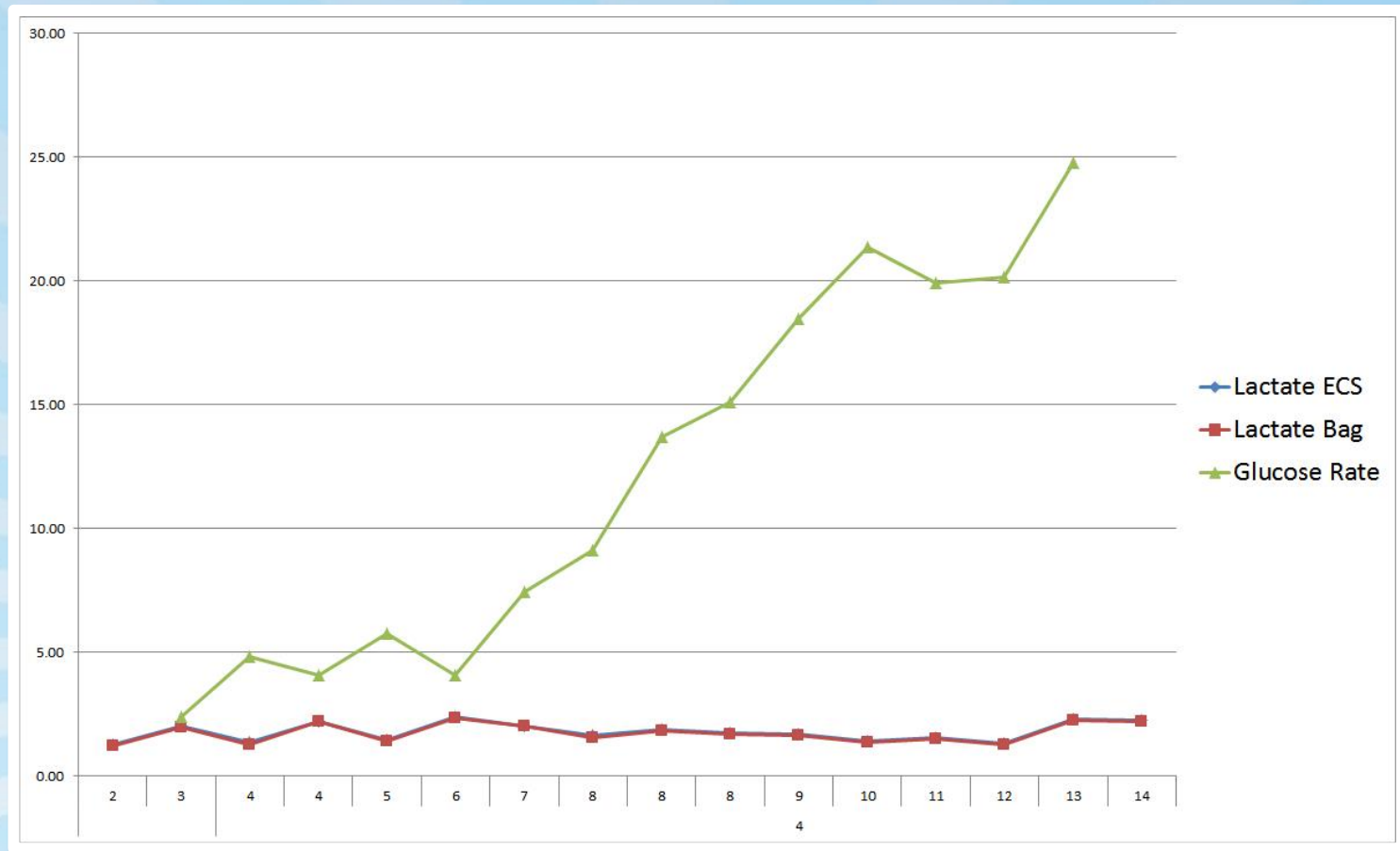
Data showing exchange across the fiber



Data showing exchange across the fiber



Data showing exchange across the fiber

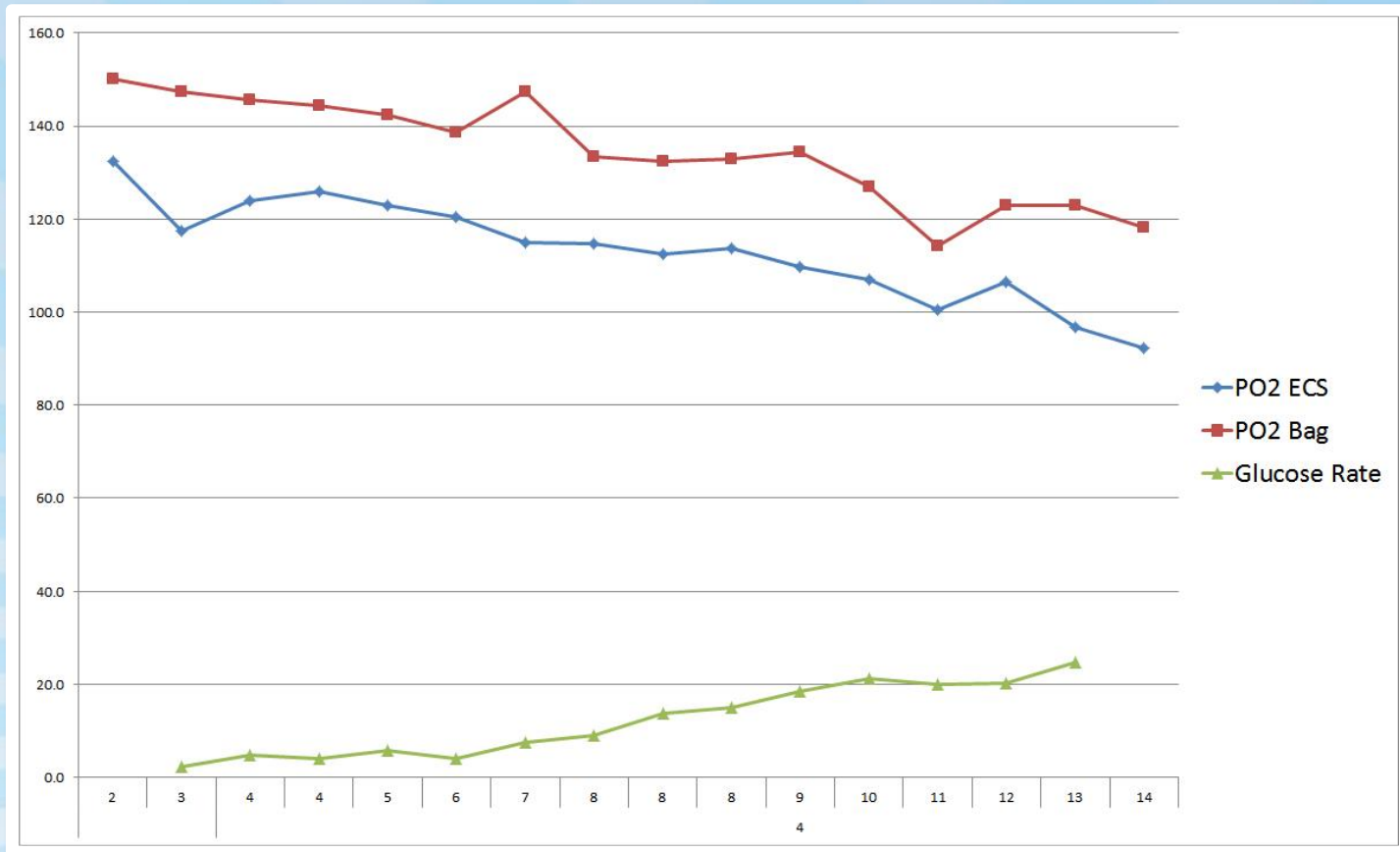


Uniformity of Cell Distribution in Cartridge



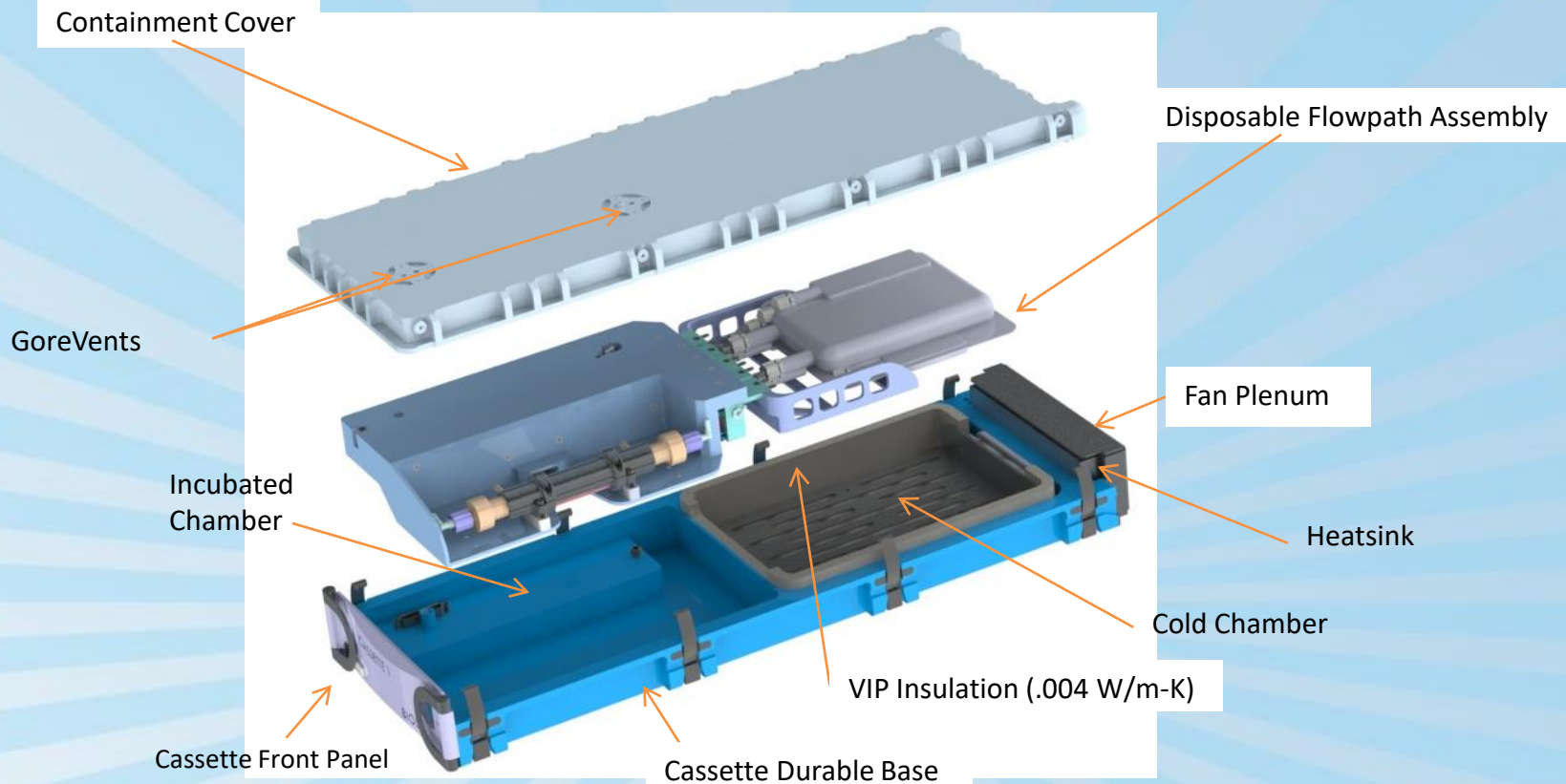


Data showing exchange across the fiber





Cassette Assembly



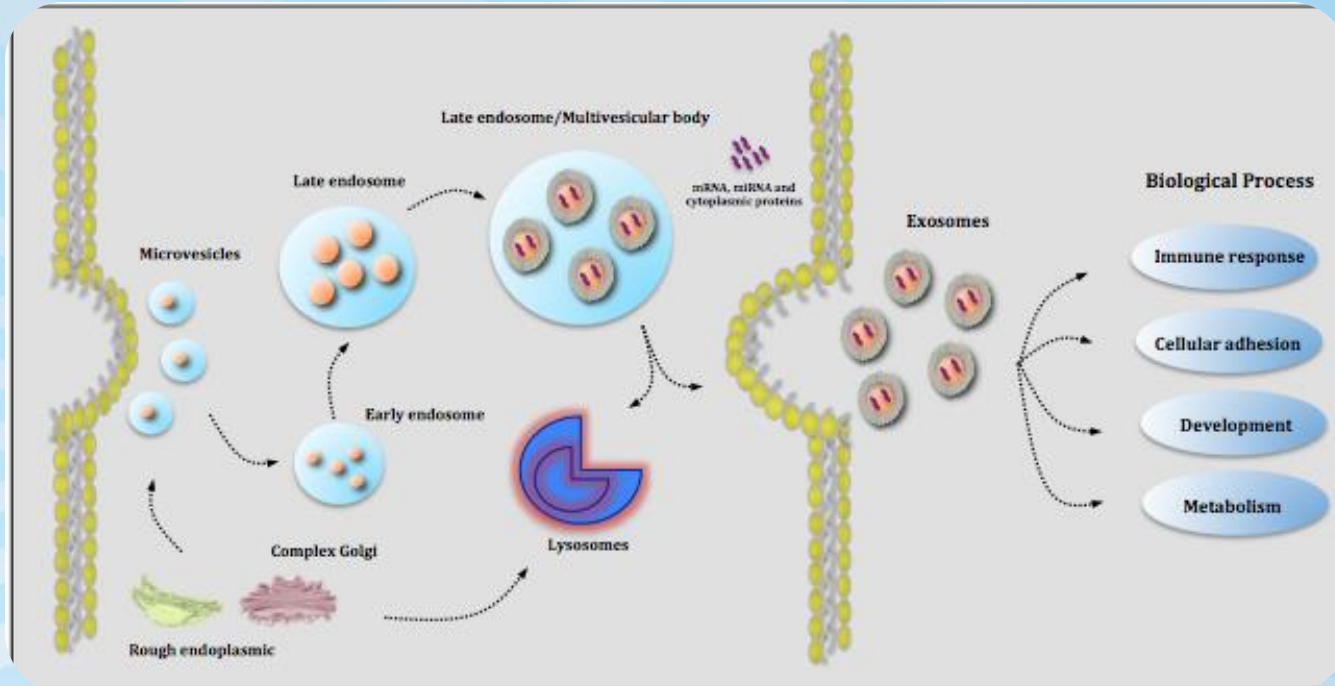
Trends in Bio-manufacturing

- Single use technology
- Perfusion/continuous production
- Higher productivity cells
- Smaller batch sizes, higher bioactivity
- Proteins becoming more complex



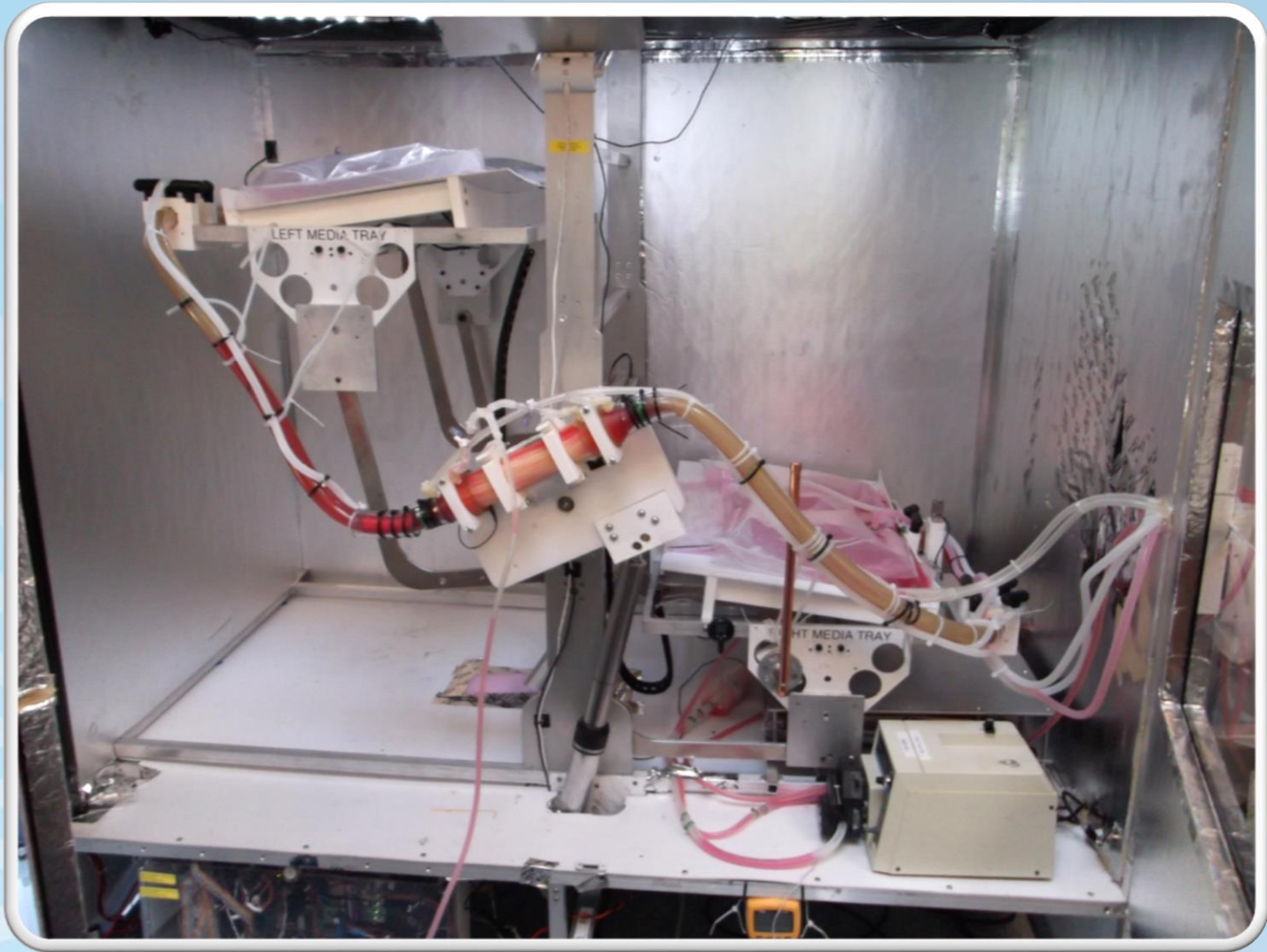
Exosomes

- Cell-derived vesicles in biological fluids
- Including medium of cultured cells
- Dia. between 30 and 100 nm
- Contain cellular proteins and RNA
- Facilitate cell-to-cell transfer of cargo
- May play a role in cell-to-cell signaling
- May mediate adaptive immune responses



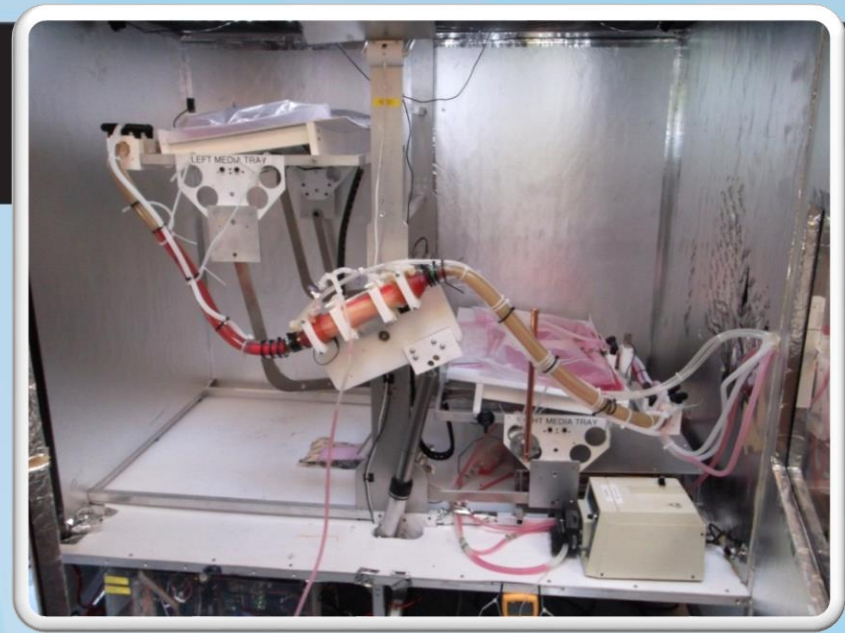
Limitations to Current Methods

- Non-Disposable. Cleaning validation
- Cells in suspension
- Surfactant and effect on KLA
- Dilute product
- Large footprint and expense
- Media, expensive, contains proteins and surfactants
- Bigger the reactor, less uniformity
- Conditions “controlled”



Advantages

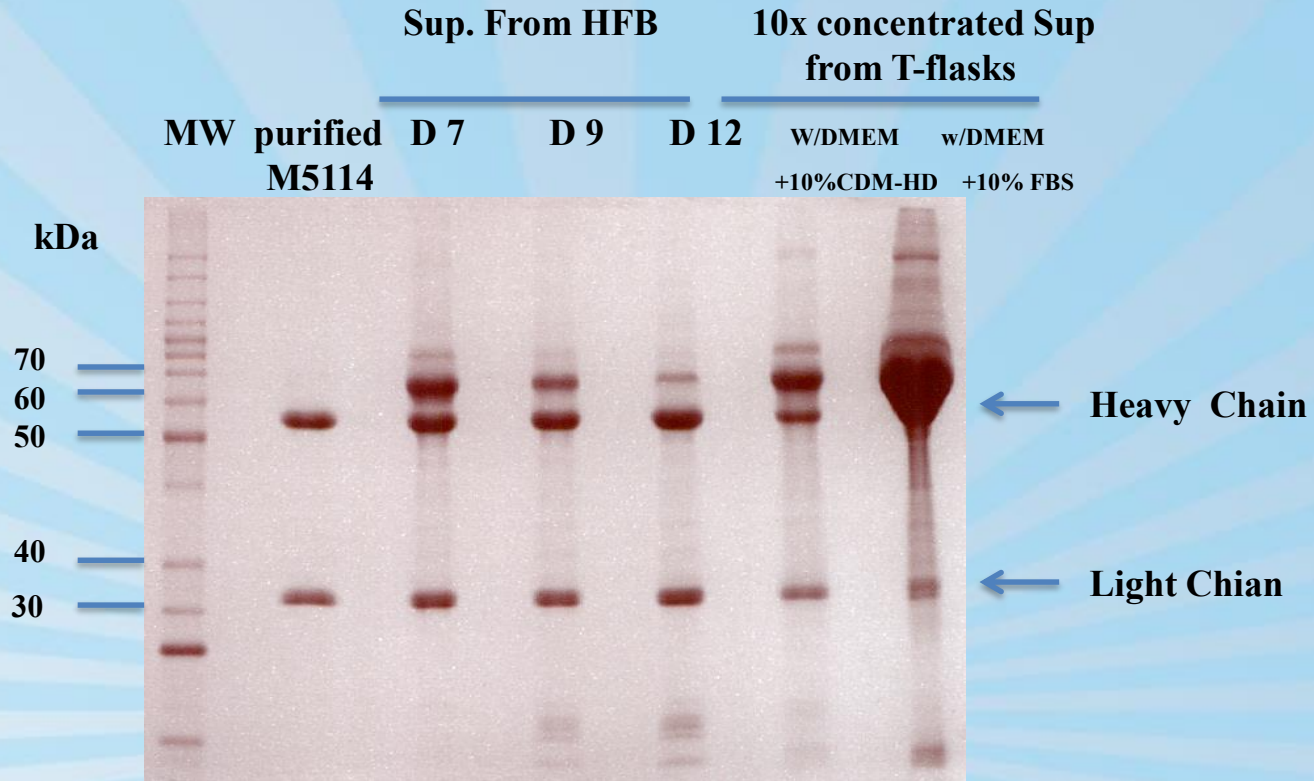
- Inexpensive medium
- Small footprint
- Disposable
- No seed reactor
- Enclosed environment
- No adaptation to suspension culture required
- Continuous harvest for labile and toxic proteins
- Small volume of reactor provides increased uniformity throughout the culture volume
- Medium is replaced, not controlled
- Especially suited for complex, highly glycosylated proteins





- Concentrated product
- Consistent continuous production over time
- Protein free medium
- Single use technology
- Can replace 500 liter to 10,000 liter stirred tanks





SDS-PAGE analysis of RAb M5114 supernatant